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(71) Applicant (for all designated States except US): **SOLEXA LIMITED** [GB/GB]; Chesterford Research Park, Little Chesterford, Nr. Saffron Walden, Essex CB10 1XL (GB).

(72) Inventors; and

(75) Inventors/Applicants (for US only): **MILTON, John** [GB/GB]; Solexa Limited, Chesterford Research Park,

Little Chesterford, Nr. Saffron Walden, Essex CB10 1XL (GB). **RUEDIGER, Silke** [DE/GB]; Solexa Limited, Chesterford Research Park, Little Chesterford, Nr. Saffron Walden, Essex CB10 1XL (GB). **LIU, Xiaohai** [CN/GB]; Solexa Limited, Chesterford Research Park, Little Chesterford, Nr. Saffron Walden, Essex CB10 1XL (GB).

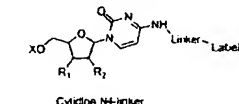
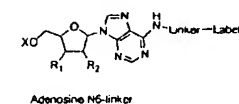
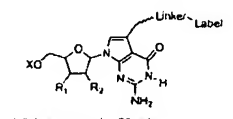
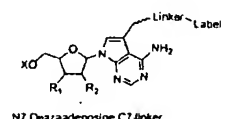
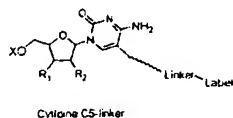
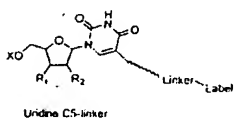
(74) Agents: **MURPHY, Colm, Damien et al.**; Boulton Wade Tennant, 70 Gray's Inn Road, London WC1X 8BT (GB).

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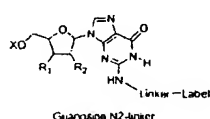
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(54) Title: LABELLED NUCLEOTIDES

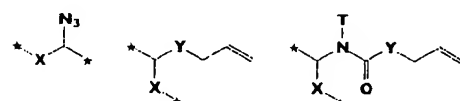


where R<sub>1</sub> and R<sub>2</sub>, which may be the same or different, are each selected from H, OH, or any group which can be transformed into an OH. Suitable groups for R<sub>1</sub> and R<sub>2</sub> are described in Figure 3

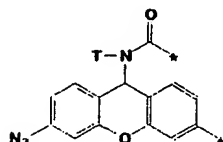
X = H, phosphate, diphosphate or triphosphate



(57) Abstract: The invention provides a nucleotide or nucleoside having a base attached to a detectable label via a cleavable linker, characterised in that the cleavable linker contains a moiety selected from the group comprising : Formula (I) (wherein X is selected from the group comprising O, S, NH and NQ wherein Q is a C<sub>1-10</sub> substituted or unsubstituted alkyl group, Y is selected from the group comprising O, S, NH and N(allyl). T is hydrogen or a C<sub>1-10</sub> substituted or unsubstituted alkyl group and \* indicates where the moiety is connected to the remainder of the nucleotide or nucleoside).



(I)





Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM),  
European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE,  
ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, RO,  
SE, SI, SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM,  
GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

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FIELD OF THE INVENTION

5           This invention relates to labelled nucleotides.  
In particular, this invention discloses nucleotides  
having a removable detectable label and their use in  
polynucleotide sequencing methods.

10    BACKGROUND

Advances in the study of molecules have been led,  
in part, by improvement in technologies used to  
characterise the molecules or their biological  
reactions. In particular, the study of the nucleic  
15   acids DNA and RNA has benefited from developing  
technologies used for sequence analysis and the study  
of hybridisation events.

An example of the technologies that have improved  
the study of nucleic acids, is the development of  
20   fabricated arrays of immobilised nucleic acids. These  
arrays consist typically of a high-density matrix of  
polynucleotides immobilised onto a solid support  
material. See, e.g., Fodor et al., *Trends Biotech.*  
12:19-26, 1994, which describes ways of assembling the  
25   nucleic acids using a chemically sensitized glass  
surface protected by a mask, but exposed at defined  
areas to allow attachment of suitably modified  
nucleotide phosphoramidites. Fabricated arrays can  
also be manufactured by the technique of "spotting"  
30   known polynucleotides onto a solid support at  
predetermined positions (e.g., Stimpson et al., *Proc.*  
*Natl. Acad. Sci. USA* 92:6379-6383, 1995).

A further development in array technology is the  
attachment of the polynucleotides to the solid support  
35   material to form single molecule arrays. Arrays of  
this type are disclosed in International Patent App.

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WO 00/06770. The advantage of these arrays is that reactions can be monitored at the single molecule level and information on large numbers of single molecules can be collated from a single reaction.

5 For DNA arrays to be useful, the sequences of the molecules must be determined. U.S. Pat. No. 5,302,509 discloses a method to sequence polynucleotides immobilised on a solid support. The method relies on the incorporation of 3'-blocked bases A, G, C and T,  
10 each of which has a distinct fluorescent label, into the immobilised polynucleotide, in the presence of DNA polymerase. The polymerase incorporates a base complementary to the target polynucleotide, but is prevented from further addition by the 3'-blocking  
15 group. The label of the incorporated base can then be determined and the blocking group removed by chemical cleavage to allow further polymerisation to occur.

Welch et al. (*Chem. Eur. J.* 5(3):951-960, 1999) describes the synthesis of nucleotide triphosphates  
20 modified with a 3'-O-blocking group that is photolabile and fluorescent. The modified nucleotides are intended for use in DNA sequencing experiments. However, these nucleotides proved to be difficult to incorporate onto an existing polynucleotide, due to an  
25 inability to fit into the polymerase enzyme active site.

Zhu et al. (*Cytometry* 28:206-211, 1997) also discloses the use of fluorescent labels attached to a nucleotide via the base group. The labelled  
30 nucleotides are intended for use in fluorescence in situ hybridisation (FISH) experiments, where a series of incorporated labelled nucleotides is required to produce a fluorescent "bar code".

WO99/57321 describes the use of nucleotides  
35 comprising fluorophores linked to the nucleotide by chemically or photochemically cleavable linker

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moieties.

WO00/53812 and EP-A2-1 291 354 disclose nucleotide compounds of general structure Fluorophore-S-S-Linker-Nucleotide and their use in nucleic acid assay methods. WO00/53812 also makes reference to periodate cleavage of a cis-glycol linkage between nucleotide and fluorophore.

WO 01/92284 discloses the use of enzyme-cleavable groups linking blocking and reporting moieties to nucleotides. It is preferred that these enzyme-cleavable groups are the same, i.e. that both the blocking and reporter moieties are attached to the nucleotide by a chain comprising a group cleavable by a common enzyme. Cleavable groups described in WO 01/92284 are esters and amides, cleavable by esterases and amidases respectively.

WO02/29003 describes nucleotides analogues that contain a label linked through cleavable linkers to the nucleotide base, or an analogue of the base. Photocleavable linkers comprising 2-nitrobenzyl moieties are described.

#### SUMMARY OF THE INVENTION

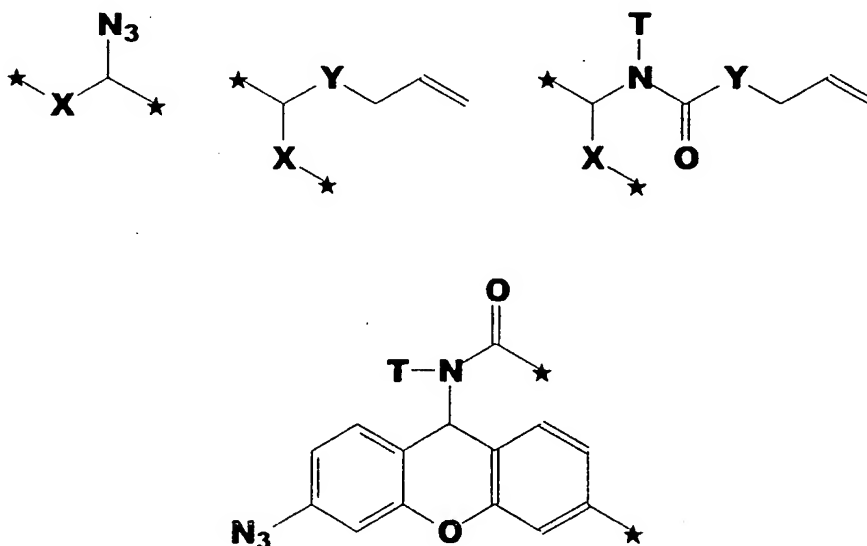
The present inventors have surprisingly found that certain linkers which connect the bases of nucleotides to detachable labels, e.g. fluorophores, may be cleaved using water-soluble phosphines or water-soluble transition metal catalysts formed from a transition metal and at least partially water-soluble ligands. In aqueous solution the latter form at least partially water-soluble transition metal complexes.

Labelled nucleotides comprising such linkers and methods for using them are clearly advantageous in the context of techniques carried out in aqueous media such as sequencing reactions, polynucleotide synthesis, nucleic acid amplification, nucleic acid

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hybridization assays, single nucleotide polymorphism studies, and other techniques using enzymes such as polymerases, reverse transcriptases, terminal transferases, or other DNA modifying enzymes. The invention is especially useful in techniques that use labelled dNTPs, such as nick translation, random primer labeling, end-labeling (e.g., with terminal deoxynucleotidyltransferase), reverse transcription, or nucleic acid amplification.

According to a first aspect of the invention, there is provided a nucleotide or nucleoside having a base attached to a detectable label via a cleavable linker, characterised in that the cleavable linker contains a moiety selected from the group comprising:

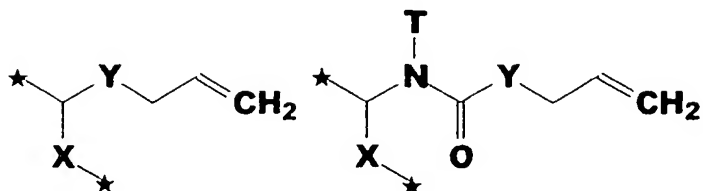


(wherein X is selected from the group comprising O, S, NH and NQ wherein Q is a C<sub>1-10</sub> substituted or unsubstituted alkyl group, Y is selected from the group comprising O, S, NH and N(allyl), T is hydrogen or a C<sub>1-10</sub> substituted or unsubstituted alkyl group and \* indicates where the moiety is connected to the remainder of the nucleotide or nucleoside).

According to a second aspect of the invention, there is provided a method of cleaving a linker that

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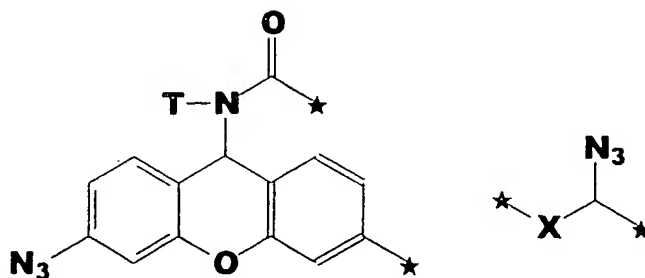
contains a moiety selected from the groups comprising:



5 (wherein X is selected from the group comprising O, S, NH and NQ wherein Q is a C<sub>1-10</sub> substituted or unsubstituted alkyl group, Y is selected from the group comprising O, S, NH and N(allyl), T is hydrogen or a C<sub>1-10</sub> substituted or unsubstituted alkyl group and  
 10 \* indicates where the moiety is connected to the remainder of a nucleotide or nucleoside), said linker being present in the nucleotide or nucleoside and connecting the base thereof to a detectable label, said method comprising contacting the nucleotide or  
 15 nucleoside with a water-soluble phosphine-based transition metal catalyst.

According to a third aspect of the invention, there is provided a method of cleaving a linker that contains a moiety selected from the groups comprising:

20



(wherein X is selected from the group comprising O, S, NH and NQ wherein Q is a C<sub>1-10</sub> substituted or  
 25 unsubstituted alkyl group, T is hydrogen or a C<sub>1-10</sub> substituted or unsubstituted alkyl group and \* indicates where the moiety is connected to the

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remainder of a nucleotide or nucleoside), said linker being present in the nucleotide or nucleoside and connecting the base thereof to a detectable label, said method comprising contacting the nucleotide or nucleoside with a water-soluble phosphine.

The method according to the second and third aspects of the invention are particularly useful in sequencing reactions. Such reactions constitute a further aspect of the invention. Viewed from this aspect, the invention provides a method for determining an identity of a nucleotide in a target single-stranded polynucleotide, comprising:

- (a) providing one or more of the nucleotides A, G, C and T or U in which each of said nucleotides has a base that is attached to a distinct detectable label via a linker, said linker being cleavable with a water-soluble phosphine; and a nascent polynucleotide complementary to the target polynucleotide, one of said provided nucleotides being suitable for incorporation into said nascent polynucleotide;
- (b) incorporating the nucleotide suitable for incorporation into said nascent polynucleotide; and
- (c) carrying out a method according to the second or third aspect of the invention.

#### DESCRIPTION OF THE DRAWINGS

Fig. 1 shows exemplary nucleotide structures useful in the invention. For each structure, X can be H, phosphate, diphosphate or triphosphate.  $R_1$  and  $R_2$  can be the same or different, and can be selected from H, OH, or any group which can be transformed into an OH.

Fig. 2 shows some functional molecules useful in the invention, which include some cleavable linkers. In these structures,  $R_1$  and  $R_2$  may be the same or different, and can be H, OH, or any group which can be



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transformed into an OH group, including a carbonyl.  
R<sub>3</sub> represents one or more substituents independently  
selected from alkyl, alkoxyl, amino or halogen groups.  
Alternatively, cleavable linkers may be constructed  
5 from any labile functionality used on the 3'-block.

Fig. 3 is a schematic illustration of some of the  
2' or 3' OH blocking groups which can be present in  
the nucleotides or nucleosides according to the  
invention.

10 Fig. 4 shows two cycles of incorporation of  
a fully functional T nucleoside triphosphate against a  
poly A template

#### DETAILED DESCRIPTION

15 The nucleotide or nucleoside molecules of the  
invention each have a base that is linked to a  
detectable label via linkers that may be cleaved by  
contact with water-soluble phosphines or water-soluble  
transition metal-containing catalysts described in  
20 greater detail hereinafter. Preferably moiety "T" is  
hydrogen. The base can be a purine, or a pyrimidine.  
The base can be a deazapurine. The molecule can have  
a ribose or deoxyribose sugar moiety. The ribose or  
deoxyribose sugar can include a protecting group  
25 attached via the 2' or 3' oxygen atom. The protecting  
group can be removed to expose a 3'-OH. The molecule  
can be a deoxyribonucleotide triphosphate. The  
detectable label can be a fluorophore.

The invention also embraces oligonucleotides  
30 which comprise one or more nucleotides of the  
invention. Preferably, at least one nucleotide of the  
invention is present at a terminal position in such  
aoligonucleotide.

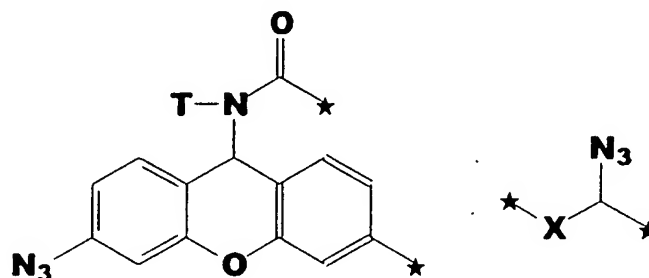
The linker may be attached to the 5-position in  
35 pyrimidines or the 7-position in purines or  
deazapurines. The characteristic feature of the

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nucleotides and nucleosides of the present invention is the amenability of the linkage to cleavage by certain water-soluble phosphines or phosphine-based transition metal catalysts. Since oligonucleotides are manipulated in aqueous solution, the advantages of this water-solubility are evident.

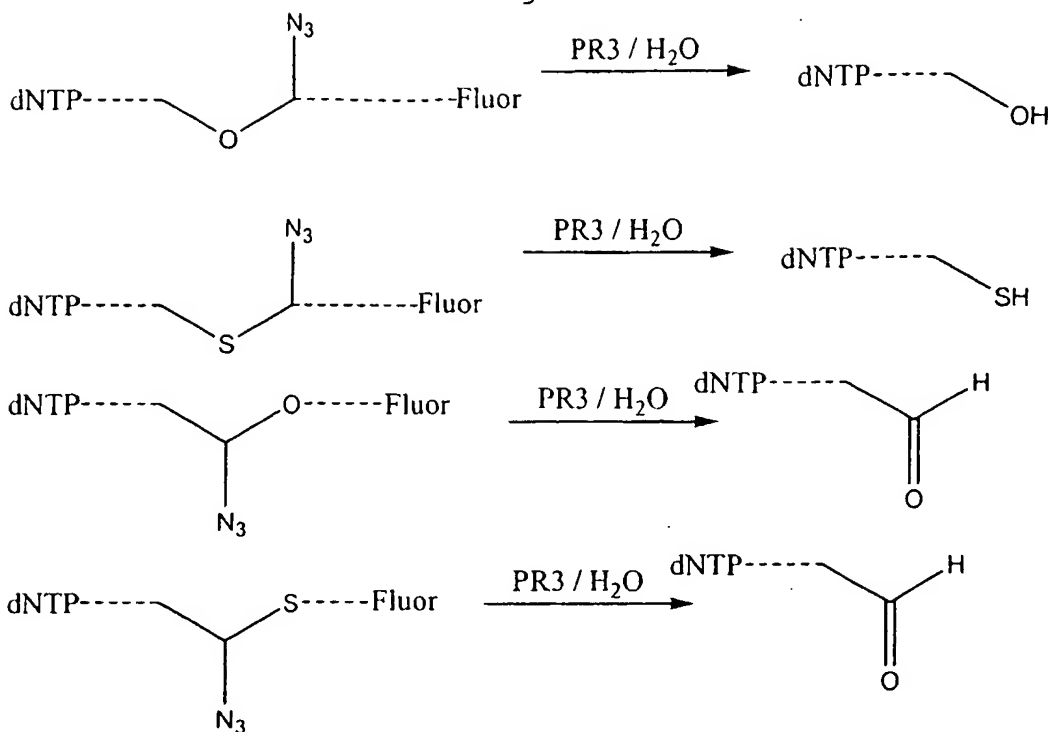
The cleavable linkages present in the nucleosides and nucleotides of the invention each comprise an allyl or azido group.

Where the linkers comprise an azide-containing group the linkers may contain a moiety of the formula:



These moieties may be present in either orientation in the linker connecting the base of the nucleotide/nucleoside with the detectable label, that is to say either of the bonds shown terminating in asterisks in each moiety may be closer to the base (or the label) in each nucleotide or nucleoside than the other asterisk shown in each structure. Thus the invention embraces nucleotides and nucleoside having schematically the following structures (shown on the left-hand side) which may be reacted with the water-soluble phosphines (described in greater detail hereinafter) to generate the products shown on the right-hand side in which the azido-containing linker has been cleaved:

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5

Whilst the connecting points \* indicate the points at which the moieties are connected to the nucleotide or nucleoside, it will be appreciated that these points are generally the points at which the moiety is connected to the remainder of the linker. In other words, the moieties described herein that contain allyl or azido groups are generally part of, rather than exclusively constitute, the cleavable linker.

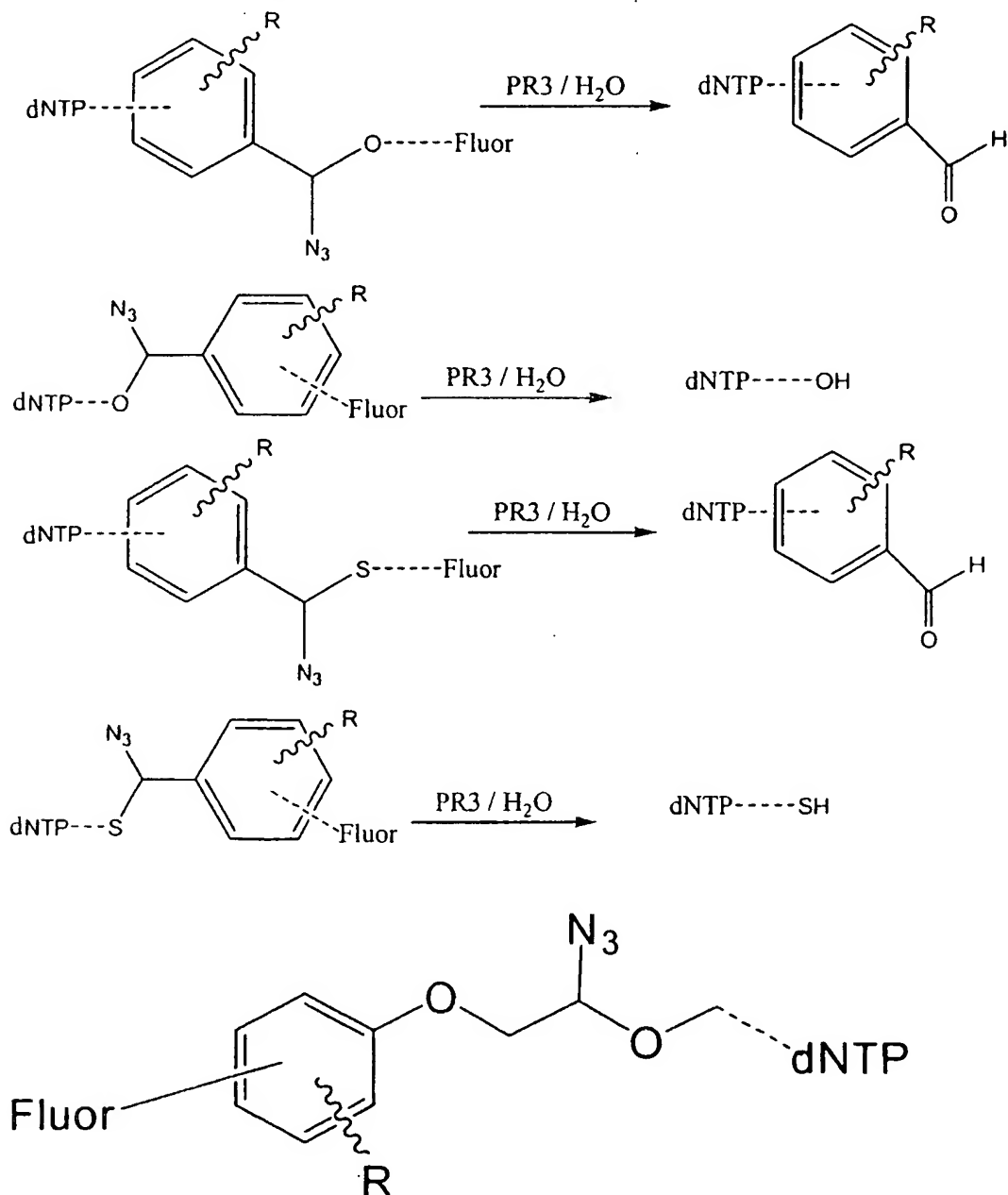
Where the moiety is of formula -X-CH(N<sub>3</sub>)-, the nature of the substituents to either side of the moiety -X-CH(N<sub>3</sub>)- affects the stability of the moiety and thus the rate at which it is cleaved. For example, where the linkage contains a substituted aryl group attached to the moiety in which the substituents (indicated as "R" in the structures immediately below) are electron-withdrawing, this is manifested in the way in which the moiety cleaves. For example, electron-withdrawing groups, serve to stabilise the linkage particularly where X is O or S. This makes

25

- 10 -

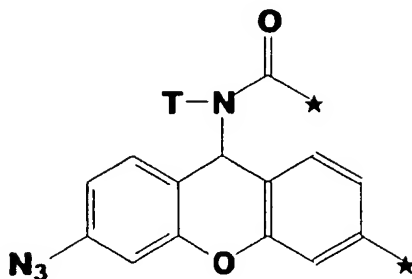
cleavage occur more slowly.

Shown below schematically are the outcomes of four cleavages of schematic nucleotide constructs and a further schematic representation of a preferred construct. The dotted lines connecting the "fluor", "dNTP" or R group to the benzene ring or the cleavable moieties indicate that substitution may be at any free position on the benzene ring and that further atoms (not shown) in the linker may be present between the cleavable motif shown and the nucleotide and detectable label which the linker connects:



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Where the azide-containing moieties are Sieber linkers, i.e. are of the formula



5

cleavage of the moiety takes place across the bond linking the central 6-membered ring of the tricycle to the amide such that a terminal amide group is left pendant to either the base or the fluorophore after cleavage.

10

It will be appreciated that the azide-containing Sieber linker moieties may contain one or more substituents which may be either electron-donating (examples include alkyl or alkoxy, e.g. C<sub>1-6</sub> alkyl or C<sub>1-6</sub> alkoxy groups) or electron-withdrawing groups (e.g. nitro, cyano, fluoro etc). Introduction of such substituents enables the skilled person to tailor the conditions under which which cleavage may be effected.

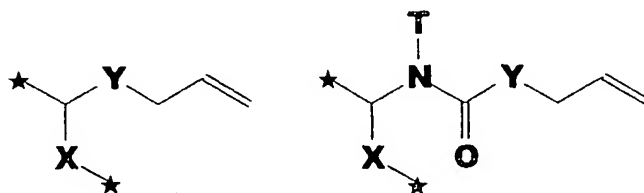
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Where the nucleotides comprise an azide group in the linker, this may be converted to the primary amine group with a thiol (in place of the phosphines), preferably a water-soluble thiol such as dithiothreitol (DTT).

20

Where the linkers comprise an allyl group, these may be of the formulae:

25



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As with the azido-containing moieties discussed above, these linking moieties may be present in either orientation in the linker connecting the base of the nucleotide/nucleoside with the detectable label.

Where the linkages comprise allyl-containing moieties, these linkers may be cleaved with water-soluble transition metal catalysts formed from a transition metal and at least partially water-soluble ligands. In aqueous solution these form at least partially water-soluble transition metal complexes.

The transition metal species serves to remove the allyl group. Where the allyl group is part of carbamate, deallylation affords the corresponding carbamic acid. This spontaneously decarboxylates to afford a hemiaminal which hydrolyses so as to effect cleavage of the linker. Corresponding chemistries operate with the analogous thiocarbamate and carbonates so as to generate compounds containing moieties of structure  $\ast\text{-C}(\text{NH}_2)\text{-X-}\ast$ . These collapse by hydrolysing, again cleaving the linker.

By aqueous solution herein is meant a liquid comprising at least 20 vol%, preferably at least 50%, for example at least 75 vol%, particularly at least 95 vol% and especially greater than above 98 vol%, ideally 100 vol% of water as the continuous phase.

Transition metals of use in forming the catalysts described herein are preferably platinum, palladium, rhodium, ruthenium, osmium and iridium. Palladium is particularly preferred.

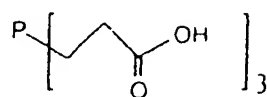
The transition metal, e.g. palladium, is conveniently introduced as a salt, e.g. as a halide. Mixed salts such as  $\text{Na}_2\text{PdCl}_4$  may also be used. Other appropriate salts and compounds will be readily determined by the skilled person and are commercially available, e.g. from Aldrich Chemical Company.

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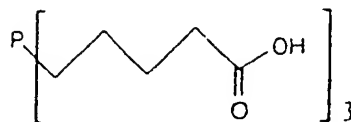
Suitable phosphines are any phosphine or mixed nitrogen-phosphine ligands known to those skilled in the art, characterised in that the ligands are derivatised so as to render them water-soluble, e.g. by introducing one or more sulfonate, amine, hydroxyl (preferably a plurality of hydroxyl) or carboxylate residues. Where amine residues are present, formation of amine salts may assist the solubilisation of the ligand and thus the metal-allyl complex. Examples of appropriate ligands are triaryl phosphines, e.g. triphenyl phosphine, derivatised so as to make them water-soluble. Also preferred are trialkyl phosphines, e.g. tri-C<sub>1-6</sub>-alkyl phosphines such as triethyl phosphines; such trialkyl phosphines are likewise derivatised so as to make them water-soluble. Sulfonate-containing and carboxylate-containing phosphines are particularly preferred; an example of the former 3,3',3''-phosphinidynetris (benzenesulfonic acid) which is commercially available from Aldrich Chemical Company as the trisodium salt; and a preferred example of the latter is tris(2-carboxyethyl)phosphine which is available from Aldrich as the hydrochloride salt. The derivatised water-soluble phosphines and nitrogen-containing phosphines described herein may be used as their salts (e.g. as the hydrochloride or sodium salts) or, for example, in the case of the sulfonic and carboxylic acid-containing phosphines described herein, as the free acids. Thus 3,3',3''-phosphinidynetris (benzenesulfonic acid) and tris(2-carboxyethyl)phosphines may be introduced either as the triacids or the trisodium salts. Other appropriate salts will be evident to those skilled in the art. The existence in salt form is not particularly important provided the phosphines are soluble in aqueous solution.

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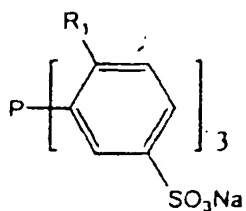
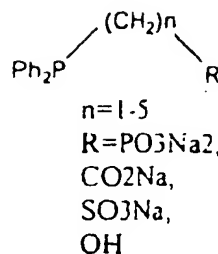
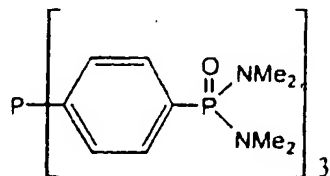
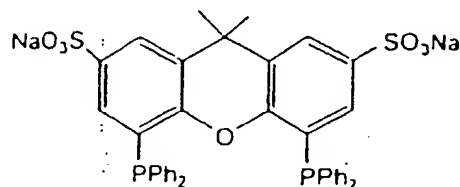
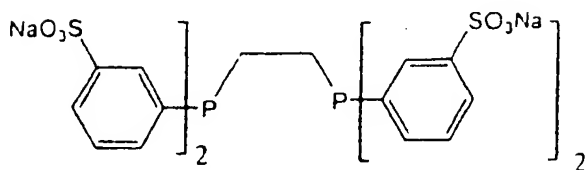
Other phosphines which may be used include the following:



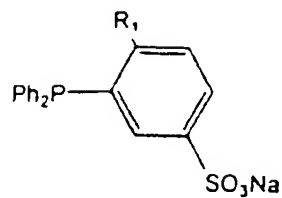
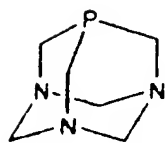
as the hydrochloride salt, neutral compound and the trisodium salt



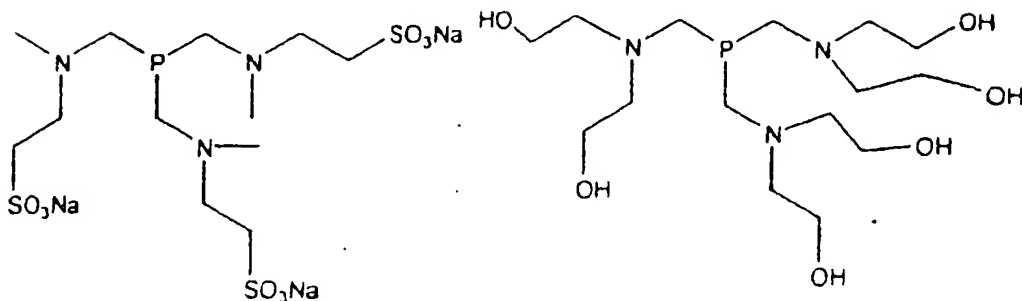
as the hydrochloride salt, neutral compound and the trisodium salt



$\text{R}_1=\text{OMe}, \text{CO}_2\text{H}, \text{CO}_2\text{Na}$



$\text{R}_1=\text{OMe}, \text{CO}_2\text{H}, \text{CO}_2\text{Na}$





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The skilled person will be aware that the atoms chelated to the transition metal in the water soluble complex may be part of mono- or polydentate ligands. Some such polydentate ligands are shown above. Whilst  
5 monodentate ligands are preferred, the invention thus also embraces methods which use water-soluble bi-, tri-, tetra-, penta- and hexadentate water-soluble phosphine and water-soluble nitrogen-containing phosphine ligands.

10 As noted earlier, the aqueous solution in which deprotection is effected need not be 100% (as the continuous phase). However, substantially pure water (e.g. at least 98 vol% and preferably about 100 vol%) is preferred. Cosolvents are generally not required.  
15 Generally, nucleotides and nucleosides are readily soluble in water (e.g. pure water) in which the linkage cleavage described herein may be effected. If desirable, one or more water-miscible cosolvents may be employed. Appropriate solvents include  
20 acetonitrile or dimethylsulfoxide, methanol, ethanol and acetone, methanol being preferred. Less preferred solvents include tetrahydrofuran (THF) and dioxane.

In the methods of cleaving allyl-containing moieties according to the invention, a soluble metal  
25 complex is formed comprising a transition metal and one or more water-soluble phosphine ligands (including water-soluble nitrogen-containing phosphine ligands). More than one type of water-soluble phosphine/nitrogen-containing phosphine ligand may be  
30 used in any given reaction although generally only one type of these classes of ligand will be used in any given catalyst. The quantity of transition metal, e.g. palladium, may be less than 1 mol% (calculated relative to the number of moles of linkage to be  
35 cleaved). Advantageously the amount of catalyst may be much less than 1 mol%, e.g. <0.50 mol%, preferably

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<0.10 mol%, particularly <0.05mol%. Even lower quantities of metal may be used, for example <0.03 or even <0.01 mol%. As those skilled in the art will be aware, however, as quantity of catalyst is reduced, so  
5 too is the speed of the reaction. The skilled person will be able to judge, in any instance, an appropriate quantity of transition metal and thus catalyst suitable for any particular reaction.

In contrast to the amount of metal required in  
10 forming the active catalyst, the quantity of water-soluble phosphine-containing ligand(s) used is preferably be greater than 1 molar equivalent (again calculated relative to the number of moles of linkage to be cleaved). Preferably greater than 4, e.g.  
15 greater than 6, for example 8-12 molar equivalents of ligand may be used. Even higher quantities of ligand e.g. >20 mole equivalents may be used if desired.

The skilled person will be able to determine the quantity of ligand best suited to any individual  
20 reaction.

Where the linkage contains an azide group, the presence of a transition metal is not necessary to effect cleavage. Thus cleavage of such linkers may be effected in the presence only of the phosphines  
25 discussed herein; these may be present in the methods of the invention as water-soluble salts, such as those discussed herein.

As is known in the art, a "nucleotide" consists of a nitrogenous base, a sugar, and one or more  
30 phosphate groups. In RNA, the sugar is a ribose, and in DNA is a deoxyribose, i.e., a sugar lacking a hydroxyl group that is present in ribose. The nitrogenous base is a derivative of purine or pyrimidine. The purines are adenosine (A) and  
35 guanidine (G), and the pyrimidines are cytidine (C) and thymidine (T) (or in the context of RNA, uracil

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(U)). The C-1 atom of deoxyribose is bonded to N-1 of a pyrimidine or N-9 of a purine. A nucleotide is also a phosphate ester of a nucleoside, with esterification occurring on the hydroxyl group attached to C-5 of the sugar. Nucleotides are usually mono, di- or triphosphates.

A "nucleoside" is structurally similar to a nucleotide, but is missing the phosphate moieties. An example of a nucleoside analog would be one in which the label is linked to the base and there is no phosphate group attached to the sugar molecule.

Although the base is usually referred to as a purine or pyrimidine, the skilled person will appreciate that derivatives and analogs are available which do not alter the capability of the nucleotide or nucleoside to undergo Watson-Crick base pairing. "Derivative" or "analog" means a compound or molecule whose core structure is the same as, or closely resembles that of, a parent compound, but which has a chemical or physical modification, such as a different or additional side group, which allows the derivative nucleotide or nucleoside to be linked to another molecule. For example, the base can be a deazapurine.

The modified nucleotides or nucleotides disclosed and claimed herein are examples of such derivatives or analogues with the addition of detectable labels connected to the bases by the cleavable allyl- or azido-containing linkers described herein. Thus the terms nucleotides and nucleosides as used herein will be understood to embrace such modified nucleosides and nucleotides.

The derivatives should be capable of undergoing Watson-Crick pairing. "Derivative" and "analogue" also mean a synthetic nucleotide or nucleoside derivative having modified base moieties and/or modified sugar moieties. Such derivatives and analogs

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are discussed in, e.g., Scheit, *Nucleotide Analogs* (John Wiley & Son, 1980) and Uhlman et al., *Chemical Reviews* 90:543-584, 1990. Nucleotide analogs can also comprise modified phosphodiester linkages, including  
5 phosphorothioate, phosphorodithioate, alkyl-phosphonate, phosphoranilidate and phosphoramidate linkages. The analogs should be capable of undergoing Watson-Crick base pairing. "Derivative", "analog" and "modified" as used herein, may be used  
10 interchangeably, and are encompassed by the terms "nucleotide" and "nucleoside" defined herein.

The methods of the present invention make use of conventional detectable labels. Detection can be carried out by any suitable method, including  
15 fluorescence spectroscopy or by other optical means. The preferred label is a fluorophore, which, after absorption of energy, emits radiation at a defined wavelength. Many suitable fluorescent labels are known. For example, Welch et al. (*Chem. Eur. J.*  
20 5(3):951-960, 1999) discloses dansyl-functionalised fluorescent moieties that can be used in the present invention. Zhu et al. (*Cytometry* 28:206-211, 1997) describes the use of the fluorescent labels Cy3 and Cy5, which can also be used in the present invention.  
25 Labels suitable for use are also disclosed in Prober et al. (*Science* 238:336-341, 1987); Connell et al. (*BioTechniques* 5(4):342-384, 1987), Ansorge et al. (*Nucl. Acids Res.* 15(11):4593-4602, 1987) and Smith et al. (*Nature* 321:674, 1986). Other commercially  
30 available fluorescent labels include, but are not limited to, fluorescein, rhodamine (including TMR, Texas red and Rox), alexa, bodipy, acridine, coumarin, pyrene, benzanthracene and the cyanins.

Multiple labels can also be used in the  
35 invention. For example, bi-fluorophore FRET cassettes (*Tet. Letts.* 46:8867-8871, 2000) are well known in the

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art and can be utilised in the present invention.  
Multi-fluor dendrimeric systems (*J. Amer. Chem. Soc.*  
123:8101-8108, 2001) can also be used.

Although fluorescent labels are preferred, other  
5 forms of detectable labels will be apparent as useful  
to those of ordinary skill. For example,  
microparticles, including quantum dots (Empodocles, et  
al., *Nature* 399:126-130, 1999), gold nanoparticles  
(Reichert et al., *Anal. Chem.* 72:6025-6029, 2000),  
10 microbeads (Lacoste et al., *Proc. Natl. Acad. Sci USA*  
97(17):9461-9466, 2000), and tags detectable by mass  
spectrometry can all be used.

Multi-component labels can also be used in the  
invention. A multi-component label is one which is  
15 dependent on the interaction with a further compound  
for detection. The most common multi-component label  
used in biology is the biotin-streptavidin system.  
Biotin is used as the label attached to the nucleotide  
base. Streptavidin is then added separately to enable  
20 detection to occur. Other multi-component systems are  
available. For example, dinitrophenol has a  
commercially available fluorescent antibody that can  
be used for detection.

The label (or label and linker construct) can be  
25 of a size or structure sufficient to act as a block to  
the incorporation of a further nucleotide onto the  
nucleotide of the invention. This permits controlled  
polymerization to be carried out. The block can be  
due to steric hindrance, or can be due to a  
30 combination of size, charge and structure.

The invention will be further described primarily  
with reference to nucleotides. However, unless  
indicated otherwise, the references herein to  
nucleotides are also intended to be applicable to  
35 nucleosides. The invention will also be further  
described with reference to DNA, although the

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description will also be applicable to RNA, PNA, and other nucleic acids, unless otherwise indicated.

The modified nucleotides of the invention use a cleavable linker to attach the label to the  
5 nucleotide. The use of a cleavable linker ensures that the label can, if required, be removed after detection, avoiding any interfering signal with any labelled nucleotide incorporated subsequently.

The use of the term "cleavable linker" is not  
10 meant to imply that the whole linker is required to be removed from the nucleotide base. The cleavage site can be located at a position on the linker that ensures that part of the linker remains attached to the nucleotide base after cleavage.

15 The linker can be attached at any position on the nucleotide base provided that Watson-Crick base pairing can still be carried out. In the context of purine bases, it is preferred if the linker is attached via the 7-position of the purine or the  
20 preferred deazapurine analogue, via an 8-modified purine, via an N-6 modified adenosine or an N-2 modified guanine. For pyrimidines, attachment is preferably via the 5-position on cytidine, thymidine or uracil and the N-4 position on cytosine. Suitable  
25 nucleotide structures are shown in Fig. 1. For each structure in Fig. 1, X can be H, phosphate, diphosphate or triphosphate.  $R_1$  and  $R_2$  can be the same or different, and can be selected from H, OH, or any group which can be transformed into an OH, including,  
30 but not limited to, a carbonyl.

Suitable linkers comprise the azide- and allyl-containing moieties discussed earlier. However, in addition to these cleavable moieties, other cleavable motifs may of course also be present in the linkers.  
35 Referring to Fig.2, examples of these include, but are not limited to, disulfide linkers (1), acid labile

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moieties (2, 3, 4 and 5; including dialkoxybenzyl  
moieties (e.g., 2), Sieber linkers (e.g., 3), indole  
moieties (e.g., 4), t-butyl Sieber moieties (e.g.,  
5)), electrophilically cleavable moieties,  
5 nucleophilically cleavable moieties, photocleavable  
moieties, cleavage under reductive conditions,  
oxidative conditions, cleavage via use of safety-catch  
moieties, and cleavage by elimination mechanisms.  
Examples of such moieties are described in  
10 WO03/048387.

As well as the moiety cleavable by water-soluble  
phosphines or transition metal-based catalysts  
described herein, the cleavable linkages may also  
comprise a spacer unit. The spacer distances the  
15 nucleotide base from the cleavage site or label. The  
length of the moiety is unimportant provided that the  
label is held a sufficient distance from the  
nucleotide so as not to interfere with any interaction  
between the nucleotide and an enzyme.

20 Such spacing groups may contain one or more  
arylene, e.g. phenylene, groups in the chain (i.e. a  
moiety -Ar- where the phenyl ring is part of the  
linker by way of its 1,4, 1,3 or 1,2-disposed carbon  
atoms). The phenyl ring may be substituted at its  
25 non-bonded position with one or more substituents such  
as alkyl, hydroxyl, alkyloxy, halide, nitro, carboxyl  
or cyano and the like, particularly electron-  
withdrawing groups, which electron-withdrawing is  
either by induction or resonance. An example of an  
30 electron-withdrawing group by resonance is nitro; a  
group which acts through induction is fluoro. The  
skilled person will be aware of other appropriate  
electron-withdrawing groups. The linkage in the R'  
group may also include moieties such a -O-, -S(O)<sub>q</sub>,  
35 wherein q is 0, 1 or 2 or NH or Nalkyl.

The modified nucleotides can also comprise

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additional groups or modifications to the sugar group. For example, a dideoxyribose derivative, lacking both hydroxyl groups on the ribose ring structure (at the 2' and 3' positions), can be prepared and used as a block to further nucleotide incorporation on a growing oligonucleotide strand. The protecting group is intended to prevent nucleotide incorporation onto a nascent polynucleotide strand, and can be removed under defined conditions to allow polymerisation to occur. In contrast to the prior art, there need be no detectable label attached at the ribose 3' position. This allows that steric hindrance with the polymerase enzyme to be reduced, while still allowing control of incorporation using the protecting group.

The skilled person will appreciate how to attach a suitable protecting group to the ribose ring to block interactions with the 3'-OH. The protecting group can be attached directly at the 3' position, or can be attached at the 2' position (the protecting group being of sufficient size or charge to block interactions at the 3' position). Alternatively, the protecting group can be attached at both the 3' and 2' positions, and can be cleaved to expose the 3'OH group.

Suitable protecting groups will be apparent to the skilled person, and can be formed from any suitable protecting group disclosed in "Protective Groups in Organic Synthesis", T.W.Greene and P.G.M. Wuts, 3rd Ed., Wiley Interscience, New York. The protecting group should be removable (or modifiable) to produce a 3' OH group. The process used to obtain the 3' OH group can be any suitable chemical or enzymic reaction.

Preferably, the blocking, or protecting group is an allyl group or a group of the structure



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wherein Z is any of  $-C(R')_2-O-R''$ ,  $-C(R')_2-N(R'')_2$ ,  $-C(R')_2-N(H)R''$ ,  $-C(R')_2-S-R''$  and  $-C(R')_2-F$ ,

wherein each  $R''$  is or is part of a removable protecting group;

5 each  $R'$  is independently a hydrogen atom, an alkyl, substituted alkyl, arylalkyl, alkenyl, alkynyl, aryl, heteroaryl, heterocyclic, acyl, cyano, alkoxy, aryloxy, heteroaryloxy or amido group; or  $(R')_2$  represents an alkylidene group of formula  $=C(R''')_2$  wherein each  $R'''$  may be the same or different and is  
10 selected from the group comprising hydrogen and halogen atoms and alkyl groups; and

wherein said molecule may be reacted to yield an intermediate in which each  $R''$  is exchanged for H or, where Z is  $-C(R')_2-F$ , the F is exchanged for OH, SH or  
15  $NH_2$ , preferably OH, which intermediate dissociates under aqueous conditions to afford a molecule with a free 3'OH;

with the proviso that where Z is  $-C(R')_2-S-R''$ ,  
20 both  $R'$  groups are not H.

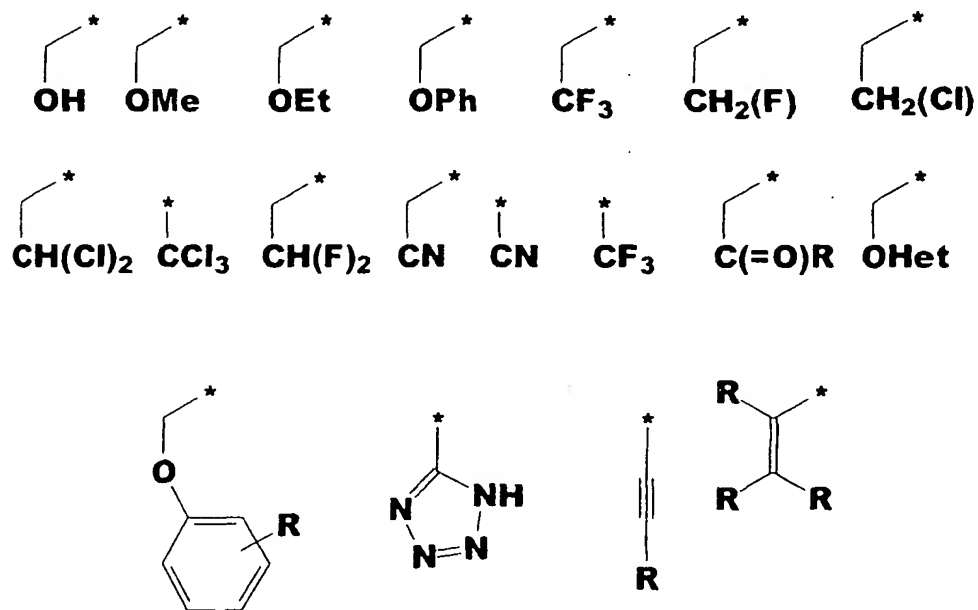
Where the blocking group is an allyl group, it may be introduced into the 3'-position using standard literature procedures such as that used by Metzker (*Nucleic Acids Research*, 22(20):4259-4267, 1994).

25 The intermediates produced advantageously spontaneously dissociate under aqueous conditions back to the natural 3' hydroxy structure, which permits further incorporation of another nucleotide. Any appropriate protecting group may be used. Preferably, Z is of formula  $-C(R')_2-O-R''$ ,  $-C(R')_2-N(R'')_2$ ,  $-C(R')_2-N(H)R''$  and  $-C(R')_2-SR''$ . Particularly preferably, Z is  
30 of the formula  $-C(R')_2-O-R''$ ,  $-C(R')_2-N(R'')_2$ , and  $-C(R')_2-SR''$ .  $R''$  may be a benzyl group or a substituted benzyl group.

35 One example of groups of structure  $-O-Z$  wherein Z

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is  $-C(R')_2-N(R'')_2$  are those in which  $-N(R'')_2$  is azido ( $-N_3$ ). One preferred such example is azidomethyl wherein each  $R'$  is H. Alternatively,  $R'$  in Z groups of formula  $-C(R')_2-N_3$  and other Z groups may be any of the other groups discussed herein. Examples of typical  $R'$  groups include  $C_{1-6}$  alkyl, particularly methyl and ethyl, and the following (in which each structure shows the bond which connects the  $R'$  moiety to the carbon atom to which it is attached in the Z groups; the asterisks (\*) indicate the points of attachment):



15

(wherein each R is an optionally substituted  $C_{1-10}$  alkyl group, an optionally substituted alkoxy group, a halogen atom or functional group such as hydroxyl, amino, cyano, nitro, carboxyl and the like) and "Het" is a heterocyclic (which may for example be a heteroaryl group). These  $R'$  groups shown above are preferred where the other  $R'$  group is the same as the first or is hydrogen. Preferred Z groups are of formula  $C(R')_2N_3$  in which the  $R'$  groups are selected

25

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from the structures given above and hydrogen; or in which  $(R')_2$  represents an alkylidene group of formula  $=C(R''')_2$ , e.g.  $=C(Me)_2$ .

Where molecules contain Z groups of formula  $C(R')_2N_3$ , the azido group may be converted to amino by contacting such molecules with the phosphine or nitrogen-containing phosphines ligands described in detail in connection with the transition metal complexes which serve to cleave the allyl groups from compounds of formula PN-O-allyl, formula R-O-allyl,  $R_2N(allyl)$ ,  $RNH(allyl)$ ,  $RN(allyl)_2$  and R-S-allyl. When transforming azido to amino, however, no transition metal is necessary. Alternatively, the azido group in Z groups of formula  $C(R')_2N_3$  may be converted to amino by contacting such molecules with the thiols, in particular water-soluble thiols such as dithiothreitol (DTT).

The labile linker may, and preferably does, consist of functionality cleavable under identical conditions to the block. This makes the deprotection process more efficient since only a single treatment will be required to cleave both the label and the block. For example, where the linkage contains an allyl moiety as discussed and claimed herein and the blocking group is an allyl group, both linkage and blocking group will be cleavable under identical conditions. Similarly, if the linkage contains an azido moiety as discussed and claimed herein and the blocking group comprises an azido moiety, e.g. is of formula Z wherein R" is  $N_3$  as discussed hereinbefore, both linkage and blocking group will be cleavable under identical conditions. The blocking group may of course be deprotected under entirely different chemical conditions to those required to cleave the linker.

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The term "alkyl" covers both straight chain and branched chain alkyl groups. Unless the context indicates otherwise, the term "alkyl" refers to groups having 1 to 10 carbon atoms, for example 1 to 8 carbon atoms, and typically from 1 to 6 carbon atoms, for example from 1 to 4 carbon atoms. Examples of alkyl groups include methyl, ethyl, propyl, isopropyl, n-butyl, isobutyl, tert-butyl, n-pentyl, 2-pentyl, 3-pentyl, 2-methyl butyl, 3-methyl butyl, and n-hexyl and its isomers.

Examples of cycloalkyl groups are those having from 3 to 10 ring atoms, particular examples including those derived from cyclopropane, cyclobutane, cyclopentane, cyclohexane and cycloheptane, bicycloheptane and decalin.

Where alkyl (including cycloalkyl) groups are substituted, particularly where these form either both of the R' groups of the molecules of the invention, examples of appropriate substituents include halogen substituents or functional groups such as hydroxyl, amino, cyano, nitro, carboxyl and the like. Such groups may also be substituents, where appropriate, of the other R' groups in the molecules of the invention.

Examples of alkenyl groups include, but are not limited to, ethenyl (vinyl), 1-propenyl, 2-propenyl (allyl), isopropenyl, butenyl, buta-1,4-dienyl, pentenyl, and hexenyl.

Examples of cycloalkenyl groups include, but are not limited to, cyclopropenyl, cyclobutenyl, cyclopentenyl, cyclopentadienyl and cyclohexenyl.

The term alkoxy refers to C<sub>1-6</sub> alkoxy unless otherwise indicated: -OR, wherein R is a C<sub>1-6</sub>alkyl group. Examples of C<sub>1-6</sub> alkoxy groups include, but are not limited to, -OMe (methoxy), -OEt (ethoxy), -O(nPr) (n-propoxy), -O(iPr) (isopropoxy), -O(nBu) (n-butoxy), -O(sBu) (sec-butoxy), -O(iBu) (isobutoxy), and -O(tBu)

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(tert-butoxy).

The term "halogen" as used herein includes fluorine, chlorine, bromine and iodine.

The nucleotide molecules of the present invention  
5 are suitable for use in many different methods where the detection of nucleotides is required.

DNA sequencing methods, such as those outlined in U.S. Pat. No. 5,302,509 can be carried out using the nucleotide.

10 Preferably the blocked and labelled modified nucleotide constructs of the nucleotide bases A, T, C and G are recognised as substrates by the same polymerase enzyme.

In the methods described herein, each of the  
15 nucleotides can be brought into contact with the target sequentially, with removal of non-incorporated nucleotides prior to addition of the next nucleotide, where detection and removal of the label and the blocking group, if present is carried out either after  
20 addition of each nucleotide, or after addition of all four nucleotides.

In the methods, all of the nucleotides can be brought into contact with the target simultaneously, i.e., a composition comprising all of the different  
25 nucleotides is brought into contact with the target, and non-incorporated nucleotides are removed prior to detection and subsequent to removal of the label and the blocking group, if present.

The four nucleotides, one of which will be  
30 complementary to the first unpaired base in the target polynucleotide, may be brought into contact with the target sequentially, optionally with removal of non-incorporated nucleotides prior to addition of the next nucleotide. Determination of the success of the  
35 incorporation may be carried out either after provision of each nucleotide, or after the addition of

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all of the nucleotides added. If it is determined after addition of fewer than four nucleotides that one has been incorporated, it is not necessary to provide further nucleotides in order to detect the nucleotides complementary to the incorporated nucleotide.

Alternatively, all of the nucleotides can be brought into contact with the target simultaneously, i.e., a composition comprising all of the different nucleotide (i.e. A, T, C and G or A, U, C and G) is brought into contact with the target, and non-incorporated nucleotides removed prior to detection and removal of the label(s). The methods involving sequential addition of nucleotides may comprise a first substep and optionally one or more subsequent substeps. In the first substep a composition comprising one, two or three of the four possible nucleotides is provided, i.e. brought into contact with, the target. Thereafter any unincorporated nucleotides may be removed and a detecting step may be conducted to determine whether one of the nucleotides has been incorporated. If one has been incorporated, the cleavage of the linker may be effected and, if necessary, a terminal amide functionality introduced thereafter to the pendant arm. In this way the identity of a nucleotide in the target polynucleotide may be determined. The nascent polynucleotide may then be extended to determine the identity of the next unpaired nucleotide in the target oligonucleotide.

If the first substep above does not lead to incorporation of a nucleotide, or if this is not known, since the presence of incorporated nucleotides is not sought immediately after the first substep, one or more subsequent substeps may be conducted in which some or all of those nucleotides not provided in the first substep are provided either, as appropriate, simultaneously or subsequently. Thereafter any

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unincorporated nucleotides may be removed and a detecting step conducted to determine whether one of the classes of nucleotide has been incorporated. If one has been incorporated, cleavage of the linker may  
5 be effected, and if necessary as an additional step or steps, terminal amide functionality introduced to the pendant arm. In this way the identity of a nucleotide in the target polynucleotide may be determined. The nascent polynucleotide may then be extended to  
10 determine the identity of the next unpaired nucleotide in the target oligonucleotide. If necessary, a third and optionally a fourth substep may be effected in a similar manner to the second substep. Obviously, once four substeps have been effected, all four possible  
15 nucleotides will have been provided and one will have been incorporated.

It is desirable to determine whether a type or class of nucleotide has been incorporated after any particular combination comprising one, two or three  
20 nucleotides has been provided. In this way the unnecessary cost and time expended in providing the other nucleotide(s) is obviated. This is not a required feature of the invention, however.

It is also desirable, where the method for  
25 sequencing comprises one or more substeps, to remove any unincorporated nucleotides before further nucleotide are provided. Again, this is not a required feature of the invention. Obviously, it is necessary that at least some and preferably as many as  
30 practicable of the unincorporated nucleotides are removed prior to the detection of the incorporated nucleotide.

A method for determining the sequence of a target polynucleotide can be carried out by contacting the  
35 target polynucleotide separately with the different nucleotides to form the complement to that of the

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target polynucleotide, and detecting the incorporation of the nucleotides. Such a method makes use of polymerisation, whereby a polymerase enzyme extends the complementary strand by incorporating the correct nucleotide complementary to that on the target. The polymerisation reaction also requires a specific primer to initiate polymerisation.

For each cycle, the incorporation of the modified nucleotide is carried out by the polymerase enzyme, and the incorporation event is then determined. Many different polymerase enzymes exist, and it will be evident to the person of ordinary skill which is most appropriate to use. Preferred enzymes include DNA polymerase I, the Klenow fragment, DNA polymerase III, T4 or T7 DNA polymerase, Taq polymerase or Vent polymerase. Polymerases engineered to have specific properties can also be used. Preferably, the molecule is incorporated by a polymerase and particularly from *Thermococcus sp.*, such as 9°N. Even more preferably, the polymerase is a mutant 9°N A485L and even more preferably is a double mutant Y409V and A485L. An example of one such preferred enzyme is *Thermococcus sp.* 9°N exo -Y409V A485L available from New England Biolabs. Examples of such appropriate polymerases are disclosed in *Proc. Natl. Acad. Sci. USA*, 1996(93), pp 5281-5285, *Nucleic Acids Research*, 1999(27), pp 2454-2553 and *Acids Research*, 2002(30), pp 605-613.

Those skilled in the art will be aware of the utility of dideoxynucleoside triphosphates in so-called Sanger sequencing methods, and related protocols (Sanger-type), which rely upon randomised chain-termination at a particular type of nucleotide. An example of a Sanger-type sequencing protocol is the BASS method described by Metzker (*infra*). Other Sanger-type sequencing methods will be known to those skilled in the art.



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Sanger and Sanger-type methods generally operate by the conducting of an experiment in which eight types of nucleotides are provided, four of which contain a 3'OH group; and four of which omit the OH group and which are labeled differently from each other. The nucleotides used which omit the 3'OH group - dideoxy nucleotides - are conventionally abbreviated to ddNTPs. As is known by the skilled person, since the ddNTPs are labeled differently, by determining the positions of the terminal nucleotides incorporated, and combining this information, the sequence of the target oligonucleotide may be determined.

It will be recognized that the nucleotides of the present invention in which the 3'OH group is either absent or blocked may be of utility in Sanger methods and related protocols since the same effect achieved by using ddNTPs may be achieved by using 3'-OH blocking groups: both prevent incorporation of subsequent nucleotides.

The use of the nucleotides according to the present invention in Sanger and Sanger-type sequencing methods form a still further aspect of this invention. Viewed from this aspect, the invention provides the use of such nucleotides in a Sanger or a Sanger-type sequencing method.

The sequencing methods are preferably carried out with the target polynucleotide arrayed on a solid support. Multiple target polynucleotides can be immobilised on the solid support through linker molecules, or can be attached to particles, e.g., microspheres, which can also be attached to a solid support material.

The polynucleotides can be attached to the solid support by a number of means, including the use of biotin-avidin interactions. Methods for immobilizing polynucleotides on a solid support are well known in

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the art, and include lithographic techniques and "spotting" individual polynucleotides in defined positions on a solid support. Suitable solid supports are known in the art, and include glass slides and beads, ceramic and silicon surfaces and plastic materials. The support is usually a flat surface although microscopic beads (microspheres) can also be used and can in turn be attached to another solid support by known means. The microspheres can be of any suitable size, typically in the range of from 10 nm to 100 nm in diameter. In a preferred embodiment, the polynucleotides are attached directly onto a planar surface, preferably a planar glass surface. Attachment will preferably be by means of a covalent linkage. Preferably, the arrays that are used are single molecule arrays that comprise polynucleotides in distinct optically resolvable areas, e.g., as disclosed in International App. No. WO 00/06770.

The sequencing method can be carried out on both single polynucleotide molecule and multi-polynucleotide molecule arrays, i.e., arrays of distinct individual polynucleotide molecules and arrays of distinct regions comprising multiple copies of one individual polynucleotide molecule. Single molecule arrays allow each individual polynucleotide to be resolved separately. The use of single molecule arrays is preferred. Sequencing single molecule arrays non-destructively allows a spatially addressable array to be formed.

The method makes use of the polymerisation reaction to generate the complementary sequence of the target. Conditions compatible with polymerization reactions will be apparent to the skilled person.

To carry out the polymerase reaction it will usually be necessary to first anneal a primer sequence

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to the target polynucleotide, the primer sequence being recognised by the polymerase enzyme and acting as an initiation site for the subsequent extension of the complementary strand. The primer sequence may be added as a separate component with respect to the target polynucleotide. Alternatively, the primer and the target polynucleotide may each be part of one single stranded molecule, with the primer portion forming an intramolecular duplex with a part of the target, i.e., a hairpin loop structure. This structure may be immobilised to the solid support at any point on the molecule. Other conditions necessary for carrying out the polymerase reaction, including temperature, pH, buffer compositions etc., will be apparent to those skilled in the art.

The modified nucleotides of the invention are then brought into contact with the target polynucleotide, to allow polymerisation to occur. The nucleotides may be added sequentially, i.e., separate addition of each nucleotide type (A, T, G or C), or added together. If they are added together, it is preferable for each nucleotide type to be labelled with a different label.

This polymerisation step is allowed to proceed for a time sufficient to allow incorporation of a nucleotide.

Nucleotides that are not incorporated are then removed, for example, by subjecting the array to a washing step, and detection of the incorporated labels may then be carried out.

Detection may be by conventional means, for example if the label is a fluorescent moiety, detection of an incorporated base may be carried out by using a confocal scanning microscope to scan the surface of the array with a laser, to image a fluorophore bound directly to the incorporated base.

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Alternatively, a sensitive 2-D detector, such as a charge-coupled detector (CCD), can be used to visualise the individual signals generated. However, other techniques such as scanning near-field optical microscopy (SNOM) are available and may be used when  
5 imaging dense arrays. For example, using SNOM, individual polynucleotides may be distinguished when separated by a distance of less than 100 nm, e.g., 10 nm to 10  $\mu$ m. For a description of scanning near-field  
10 optical microscopy, see Moyer et al., *Laser Focus World* 29:10, 1993. Suitable apparatus used for imaging polynucleotide arrays are known and the technical set-up will be apparent to the skilled person.

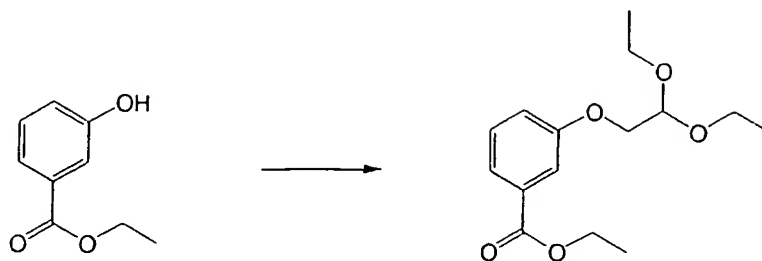
15 After detection, the label may be removed using suitable conditions that cleave the linker.

The use of the modified nucleotides is not limited to DNA sequencing techniques, and other techniques, including polynucleotide synthesis, DNA  
20 hybridisation assays and single nucleotide polymorphism studies, may also be carried out using nucleotides of the invention. Any technique that involves the interaction between a nucleotide and an enzyme may make use of the molecules of the invention.  
25 For example, the molecule may be used as a substrate for a reverse transcriptase or terminal transferase enzyme.

Suitable nucleotide structures are described in the following non-limiting Examples and are shown in  
30 the accompanying drawings.

- 35 -

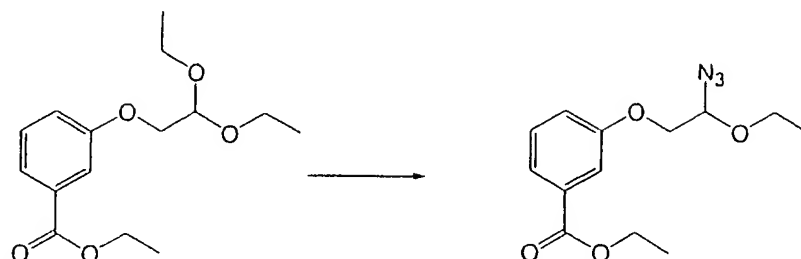
## 3-(2,2-Diethoxy-ethoxy)-benzoic acid ethyl ester



- 5 2-Bromoacetaldehyde diethyl acetal (3 ml, 20 mmol), ethyl-3-hydroxy-benzoate (1.66g, 10mmol), potassium carbonate (2.76 g, 20 mmol) and sodium iodide (0.298 g, 2 mmol) were heated at 120 °C in dimethyl formamide (DMF) (15 ml) for 17hrs. Another batch of 2-
- 10 bromoacetaldehyde diethyl acetal (3 ml, 20 mmol) was added and the reaction mixture was heated at 120 °C for another 24hrs. The reaction was cooled to room temperature and all the solvents were evaporated under reduced pressure. The residues were partitioned
- 15 between dichloromethane (DCM) (200 ml) and water (200 ml). The DCM layer was separated and the aqueous layer was back-extracted with DCM (2 x 100 ml). All the DCM extracts were combined, dried over MgSO<sub>4</sub> and evaporated under reduced pressure. The residue was
- 20 purified by a column chromatography (4 x 20 cm). The product was eluted with 100% DCM and the title compound was obtained as a colourless oil (2.21 g, 78.3%).
- 25 <sup>1</sup>HNMR [CDCl<sub>3</sub>]: 7.58 (1H, Ar-H, d J 7.7), 7.51 (1H, Ar-H, dd, J 2.4 and 1.5), 7.27 (1H, Ar-H, t, J 8.0), 7.05 (1H, Ar-H, dd, J 7.9 and 2.3), 4.79 (1H, CH, t, J 5.2), 4.20 (2H, OCH<sub>2</sub>, q, J 7.2), 3.99 (2H, ArOCH<sub>2</sub>, d, J 5.2), 3.75-3.65 (2H, OCH<sub>2</sub>, m), 3.63-3.53 (2H, OCH<sub>2</sub>, m),
- 30 1.33 (3H, CH<sub>3</sub>, t, J 7.2) and 1.19 (6H, CH<sub>3</sub>, t, J 7.1).

- 36 -

## 3-(2-Azido-2-ethoxy-ethoxy)-benzoic acid ethyl ester



5

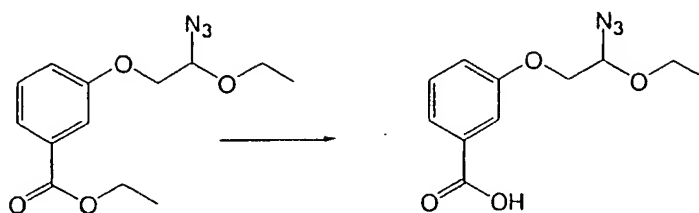
To a mixture of 3-(2,2-diethoxy-ethoxy)-benzoic acid ethyl ester (1.128 g, 4 mmol) and azidotrimethylsilane (0.584 ml, 4.4 mmol) was added  $\text{SnCl}_4$  (40  $\mu\text{l}$ ) at room temperature. After 1 hr, the precipitates were  
 10 filtered off and the filtrate was evaporated under reduced pressure. The residue was dissolved in methanol, after 10 minutes, solvent was removed under reduced pressure. The residue was purified by a column chromatography (3 x 20 cm). The product was  
 15 eluted with 10% petroleum ether (60-80 °C) in DCM. The title compound was obtained as a colourless oil (0.909 g, 81.5%).

$^1\text{H NMR}$  [ $\text{CDCl}_3$ ]: 7.51 (1H, Ar-H, d  $J$  7.7), 7.41 (1H, Ar-H, dd,  $J$  2.6 and 1.5), 7.19 (1H, Ar-H, t,  $J$  7.9), 6.96 (1H, Ar-H, ddd,  $J$  8.2, 2.7 and 0.9), 4.63 (1H,  $\text{CHN}_3$ , t,  $J$  5.1), 4.20 (2H,  $\text{OCH}_2$ , q,  $J$  7.2), 4.05-3.90 (2H,  $\text{ArOCH}_2$ , m), 3.80-3.78 (1H,  $\text{OCH}_2$ ,  $\text{H}_a$ , m), 3.55-3.47 (1H,  $\text{OCH}_2$ ,  $\text{H}_b$ , m), 1.22 (3H,  $\text{CH}_3$ , t,  $J$  7.1) and 1.13 (3H,  $\text{CH}_3$ , t,  $J$  7.1).  
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## 3-(2-Azido-2-ethoxy-ethoxy)-benzoic acid



5

3-(2,2-Diethoxy-ethoxy)-benzoic acid ethyl ester (0.279 g, 1 mmol) was stirred with 4 M aqueous sodium hydroxide (2.5 ml, 10 mmol) and ethanol (2.5 ml) at room temperature. After 4 hrs, all the solvents were removed under reduced pressure and the residue was dissolved in 50 ml water. The solution was acidified with 1 N HCl to pH 2 and then extracted with DCM (3 x 50 ml). All the DCM extracts were combined, dried over MgSO<sub>4</sub> and evaporated under reduced pressure. The title compound was obtained as a colourless solid (0.247 g, 98.4%).

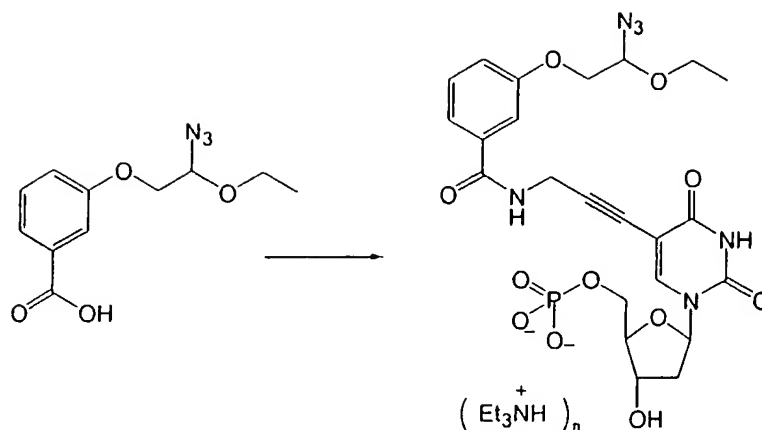
<sup>1</sup>HNMR [CDCl<sub>3</sub>]: 7.68 (1H, Ar-H, d *J* 7.7), 7.58 (1H, Ar-H, dd, *J* 2.5 and 1.5), 7.34 (1H, Ar-H, t, *J* 8.0), 7.13 (1H, Ar-H, dd, *J* 7.4 and 2.7), 4.74 (1H, CHN<sub>3</sub>, t, *J* 5.0), 4.20-4.02 (2H, ArOCH<sub>2</sub>, m), 3.95-3.80 (1H, OCH<sub>2</sub>, H<sub>a</sub>, m), 3.70-3.55 (1H, OCH<sub>2</sub>, H<sub>b</sub>, m) and 1.24 (3H, CH<sub>3</sub>, t, *J* 7.1).

25

Triethyl ammonium salt of phosphoric acid mono-[5-(5-{3-[3-(2-azido-2-ethoxy-ethoxy)-benzoylamino-prop-1-ynyl]-2,4-dioxo-3,4-dihydro-2H-pyrimidin-1-yl)-3-hydroxy-tetrahydro-furan-2-yl] ester

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- 38 -



3-(2,2-Diethoxy-ethoxy)-benzoic acid (3.77 mg, 15  $\mu$ mol) was stirred with *N,N'*-disuccinimidyl carbonate (3.84 mg, 15  $\mu$ mol) and 4-dimethylaminopyridine (DMAP) (1.83 mg, 15  $\mu$ mol) in dry DMF (1 ml) at room temperature. After 15 minutes, all the reaction mixture was added to a solution of the triethyl ammonium salt of phosphoric acid mono-{5-[5-(3-amino-prop-1-ynyl)-2,4-dioxo-3,4-dihydro-2*H*-pyrimidin-1-yl]-3-hydroxy-tetrahydro-furan-2-yl} ester (5  $\mu$ mol) in 0.1 M carbonate buffer (0.1 M NaHCO<sub>3</sub>/0.1 M Na<sub>2</sub>CO<sub>3</sub>, v/v, 1:1) (0.5 ml). After 5 hrs at room temperature, the reaction was diluted with 0.05 M triethylammonium bicarbonate (TEAB) buffer (TEAB, pH 7.5) (10 ml). The resulting solution was applied onto a short column of DEAE A-25 (1 x 5 cm). The column was initially eluted with 0.1 M TEAB buffer (50 ml) and then 0.7 M buffer (50 ml). The 0.7 M TEAB eluents were collected and evaporated under reduced pressure. The residue was co-evaporated with MeOH (2 x 10 ml) and then purified by preparative HPLC [HPLC gradient: A, 100% 0.1 M TEAB; B 100% MeCN; 0-2 min, 5% B (flow 2-10 ml/min); 2-19 min, 5-40% B (flow 10 ml/min); 19-21 min, 40-95% B (flow 10 ml/min); 21-24 min, 95% B (flow 10 ml/min); 24-26 min, 95-5% B (flow 10 ml/min); 26-30 min, 5% B (flow 10-2 ml/min)]. The product with retention time of 19.6 min was collected and evaporated under reduced



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pressure and the residue was co-evaporated with methanol (3 x 5 ml) to give the title compound as triethyl ammonium salt (3.67 mg, yield 80%). <sup>1</sup>HNMR in D<sub>2</sub>O indicated approximately 3.2 triethylammonium count ions.

<sup>1</sup>HNMR [D<sub>2</sub>O]: 7.89 (1H, H-6, s), 7.44-7.31 (3H, Ar-H, m), 7.28 (1H, Ar-H, m), 7.12 (1H, Ar-H, dt, *J* 7.1 and 2.3), 6.17 (1H, H-1', t, *J* 6.9), 4.96 (1H, CHN<sub>3</sub>, t, *J* 4.4), 4.43-4.35 (1H, H-3', m), 4.25 (2H, CHN, s), 4.18-4.06 (2H, H-5', m), 4.05-3.90 (1H, H-4', m), 3.89-3.50 (4H, ArOCH<sub>2</sub>, OCH<sub>2</sub>, m), 3.03 (19H, CH<sub>2</sub>N, q, *J* 7.3), 2.25-2.15 (2H, CH<sub>2</sub>, m), 1.13 (32H, CH<sub>3</sub>, t, *J* 7.3). <sup>31</sup>P [D<sub>2</sub>O]: 5.17 (s). MS-ES(-), *m/z* 593 [M-1].

#### 2-(1-Azido-ethoxy)-ethanol



To a mixture of 2-methyl dioxolane (0.897 ml, 10 mmol) and azidotrimethylsilane (1.6 ml, 12 mmol) was added SnCl<sub>4</sub> (40 μl) at -78 °C. After addition, the cooling bath was removed and the reaction was warmed up to room temperature. After 1 hr, the reaction was worked up by partitioning between DCM (50 ml) and saturated aqueous NaHCO<sub>3</sub> (50 ml). The aqueous layer was further extracted with DCM (20 ml). All the DCM extracts were combined, dried over MgSO<sub>4</sub> and evaporated under reduced pressure. The residue was dissolved in 10% aqueous methanol. After 2 hr at room temperature, all the solvents were removed under reduced pressure. The title compound was obtained as a colourless oil (224 mg, 17.1%).

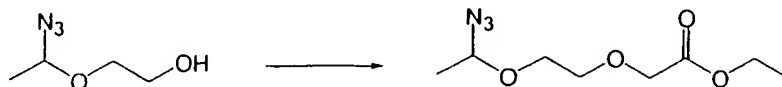
<sup>1</sup>HNMR [CDCl<sub>3</sub>]: 4.46 (1H, CHN<sub>3</sub>, q, *J* 5.7), 3.77-3.68

- 40 -

(1H, OCH<sub>2</sub>, H<sub>a</sub>, m), 3.63 (2H, OCH<sub>2</sub>, t, *J* 4.3), 3.48-3.40 (1H, OCH<sub>2</sub>, H<sub>b</sub>, m) and 1.34 (3H, CH<sub>3</sub>, d, *J* 5.7).

**[2-(1-Azido-ethoxy)-ethoxy]-acetic acid ethyl ester**

5

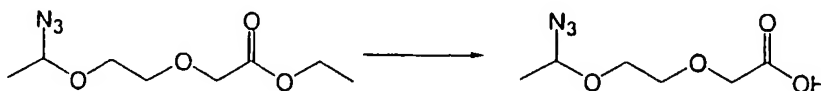


To a solution of 2-(1-azido-ethoxy)-ethanol (0.15 g, 1.14 mmol) in dry tetrahydrofuran (THF) (5 ml) was added NaH (60% dispersion, 0.08 g, 2 mmol) at 0 °C. After 15 minutes, ethyl-2-bromoacetate (0.222 ml, 2 mmol) was added. The reaction was maintained at this temperature for 15 minutes and then warmed up to room temperature. The reaction was quenched by addition of saturated aqueous NaHCO<sub>3</sub> (5 ml) after 1 hr. After a further period of 5 min, the mixture was partitioned between DCM (50 ml) and saturated aqueous NaHCO<sub>3</sub> (50 ml). The aqueous layer was further extracted with DCM (50 ml). All the DCM extracts were combined, dried over MgSO<sub>4</sub> and evaporated under reduced pressure. The title compound was obtained as an oil (0.162 g, 65.5%).

<sup>1</sup>HNMR [CDCl<sub>3</sub>]: 4.51 (1H, CHN<sub>3</sub>, q, *J* 5.6), 4.10 (2H, OCH<sub>2</sub>, q, *J* 7.1), 4.03 (2H, OCH<sub>2</sub>C(O), s), 3.85-3.77 (1H, OCH<sub>2</sub>, H<sub>a</sub>, m), 3.67-3.57 (3H, OCH<sub>2</sub>, H<sub>b</sub>, OCH<sub>2</sub>, m), 1.37 (3H, CH<sub>3</sub>, d, *J* 5.7) and 1.17 (3H, CH<sub>3</sub>, t, *J* 7.1).

**[2-(1-Azido-ethoxy)-ethoxy]-acetic acid**

30



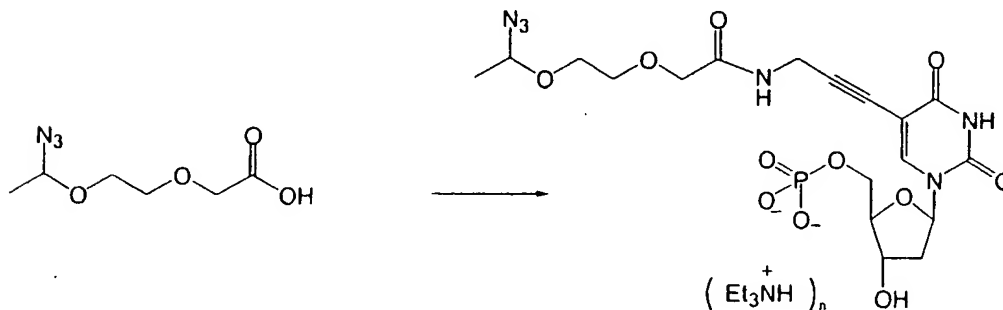
**[2-(1-Azido-ethoxy)-ethoxy]-acetic acid ethyl ester**

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(0.10 g, 0.46 mmol) was stirred with 4 M aqueous sodium hydroxide (1.15 ml, 4.6 mmol) and ethanol (1.15 ml) at room temperature. After 4 hrs, all the solvents were removed under reduced pressure and the residue was dissolved in 5 ml water. The solution was adjusted to pH 5 with 1 N  $\text{KH}_2\text{PO}_4$  and then extracted with DCM (2 x 15 ml). The aqueous layer was then acidified with 1 N HCl to pH 2 and then extracted with DCM (3 x 25 ml). All the DCM extracts were combined, dried over  $\text{MgSO}_4$  and evaporated under reduced pressure. The title compound was obtained as a colourless solid (42 mg, 48.3%).

$^1\text{H}$ NMR [ $\text{CDCl}_3$ ]: 4.55 (1H,  $\text{CHN}_3$ , q,  $J$  5.7), 4.15 (2H,  $\text{OCH}_2\text{C}(\text{O})$ , s), 3.92-3.84 (1H,  $\text{OCH}_2$ ,  $\text{H}_a$ , m), 3.74-3.70 (2H,  $\text{OCH}_2$ , m), 3.68-3.57 (1H,  $\text{OCH}_2$ ,  $\text{H}_b$ , m) and 1.44 (3H,  $\text{CH}_3$ , d,  $J$  5.7).

Triethyl ammonium salt of phosphoric acid mono-{5-[5-(3-{2-[2-(1-azido-ethoxy)-ethoxy]-acetylamino}-prop-1-ynyl)-2,4-dioxo-3,4-dihydro-2H-pyrimidin-1-yl]-3-hydroxy-tetrahydro-furan-2-yl} ester



25

[2-(1-Azido-ethoxy)-ethoxy]-acetic acid (5.7 mg, 30  $\mu\text{mol}$ ) was stirred with  $N,N'$ -disuccinimidyl carbonate (7.68 mg, 30  $\mu\text{mol}$ ) and DMAP (3.7 mg, 30  $\mu\text{mol}$ ) in dry DMF (2 ml) at room temperature. After 10 minutes, all the reaction mixture was added to a solution of the

30

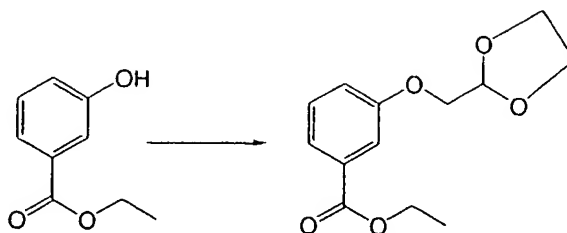
- 42 -

triethyl ammonium salt of phosphoric acid mono-{5-[5-(3-amino-prop-1-ynyl)-2,4-dioxo-3,4-dihydro-2H-pyrimidin-1-yl]-3-hydroxy-tetrahydro-furan-2-yl} ester (10  $\mu$ mol) in 0.1 M TEAB (0.5 ml). After 5 hrs at room temperature, the reaction was diluted with 0.05 M triethylammonium bicarbonate buffer (TEAB, pH 7.5) (10 ml). The resulting solution was applied onto a short column of DEAE A-25 (1 x 10 cm). The column was initially eluted with 0.1 M TEAB buffer (50 ml) and then 0.5 M buffer (50 ml). The 0.5 M TEAB eluents were collected and evaporated under reduced pressure. The residue was co-evaporated with MeOH (2 x 10 ml) and then purified by preparative HPLC [HPLC gradient: A, 100% 0.1 M TEAB; B 100% MeCN; 0-2 min, 5% B (flow 2-10 ml/min); 2-15 min, 5-15% B (flow 10 ml/min); 15-28 min, 15-22% B (flow 10 ml/min); 28-30 min, 22-95% B (flow 10 ml/min); 30-34 min, 95% B (flow 10 ml/min); 34-36 min, 95-5% B (flow 10 ml/min); 36-40 min, 5% B (flow 10-2 ml/min)]. The product with retention time of 21.3 min was collected and evaporated under reduced pressure and the residue was co-evaporated with methanol (3 x 5 ml) to give the title compound as triethyl ammonium salt (9.3 mmol, quantification at  $\lambda_{288}$  in 0.1 M TEAB buffer, 93%).  $^1\text{HNMR}$  in  $\text{D}_2\text{O}$  indicated approximately five triethylammonium count ions.

$^1\text{HNMR}$  [ $\text{D}_2\text{O}$ ]: 7.85 (1H, H-6, s), 6.15 (1H, H-1', t,  $J$  6.8), 4.75 (1H,  $\text{CHN}_3$ , q,  $J$  5.7), 4.38 (1H, H-3', m), 4.09 (2H,  $\text{OCH}_2\text{C}(\text{O})$ , s), 3.99 (2H,  $\text{CHN}$ , s), 3.95 (1H, H-4', m), 3.86-3.60 (6H,  $\text{OCH}_2\text{CH}_2\text{O}$ , H-5', m), 3.03 (30H,  $\text{CH}_2\text{N}$ , q,  $J$  7.2), 2.22-2.12 (2H,  $\text{CH}_2$ , m), 1.31 (3H,  $\text{CH}_3$ , d,  $J$  5.7) and 1.11 (45H,  $\text{CH}_3$ , t,  $J$  7.2).  $^{31}\text{P}$  [ $\text{D}_2\text{O}$ ]: 5.14 (s). MS-ES(-),  $m/z$ , 531 [M-1].

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## 3-([1,3]Dioxolan-2-ylmethoxy)-benzoic acid ethyl ester

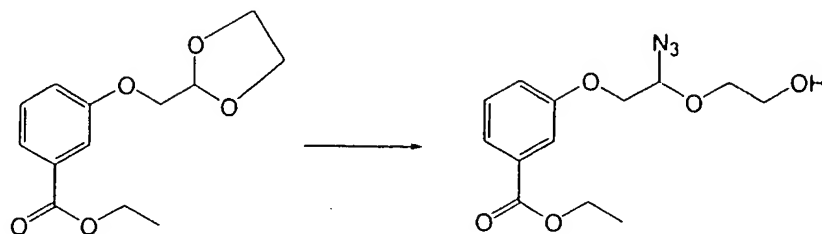


5 2-Bromomethyl-1, 3-dioxolane (8.3 ml, 80 mmol), ethyl-  
 3-hydroxy-benzoate (3.32g, 20mmol), potassium  
 carbonate (5.53 g, 40 mmol) and sodium iodide (1.2 g,  
 8 mmol) were heated at 120 °C in DMF (8 ml) for 17hrs.  
 The reaction was cooled to room temperature and all  
 10 the solvents were evaporated under reduced pressure.  
 The residues were partitioned between DCM (250 ml) and  
 water (250 ml). The DCM layer was separated and the  
 aqueous layer was back-extracted with DCM (2 x 100  
 ml). All the DCM extracts were combined, dried over  
 15  $\text{MgSO}_4$  and evaporated under reduced pressure. The  
 residue was purified by column chromatography (4 x 25  
 cm). The product was eluted with 20% petroleum ether  
 (60-80 °C) in DCM and the title compound was obtained  
 as a slightly brown oil (4.63g, 91.8%). APCI-MS, m/z  
 20 252.95 (M+1).

$^1\text{H}$ NMR [ $\text{CDCl}_3$ ]: 1.39 (3H,  $\text{CH}_3$ , t,  $J$  7.2), 3.96-4.09  
 (6H,  $\text{OCH}_2\text{CH}_2\text{O}$ ,  $\text{ArOCH}_2$ , m), 4.36 (2H,  $\text{OCH}_2$ , q,  $J$  7.2),  
 5.31 (1H, CH, t,  $J$  4.0), 7.14 (1H, Ar-H, ddd,  $J$  1.6,  
 2.6 and 8.2), 7.34 (1H, Ar-H, t,  $J$  7.9), 7.59 (1H, Ar-  
 25 H, dd,  $J$  1.5 and 2.5) and 7.67 (1H, Ar-H, dt,  $J$  1.4  
 and 7.6).

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3-[2-Azido-2-(2-hydroxy-ethoxy)-ethoxy]-benzoic acid  
ethyl ester



5

To a mixture of 3-([1, 3]-dioxolan-2-ylmethoxy)-benzoic acid ethyl ester (2.02 g, 8 mmol) and azidotrimethylsilane (1.17 ml, 8.8 mmol) was added  $\text{SnCl}_4$  (60  $\mu\text{l}$ ) at room temperature under nitrogen.

10 After 2 hr, 2% aqueous methanol (10 ml) was added to the reaction mixture and the reaction was stirred at room temperature for 30 minutes. All the solvents were evaporated under reduced pressure. The residue was co-evaporated with ethanol (2 x 10 ml). The  
15 residue was purified by a column chromatography (3 x 20 cm). The product was eluted with 0 to 1% methanol in DCM. The title compound was obtained as a colourless oil (2.01 g, 85.1%). APCI-MS,  $m/z$  267.90 ( $M-\text{N}_2+1$ ).

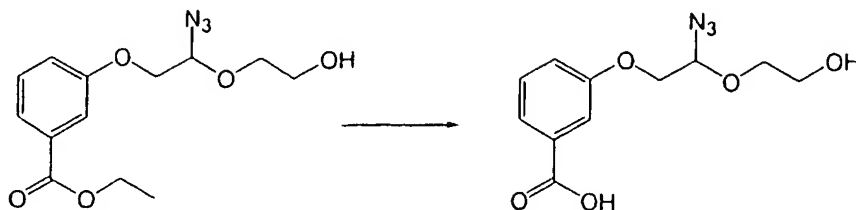
20

$^1\text{H}$ NMR [ $\text{CDCl}_3$ ]: 1.38 (3H,  $\text{CH}_3$ , t,  $J$  7.1), 3.73-3.86 (3H,  $\text{OCH}_2$ ,  $\text{H}_a$ ,  $\text{OCH}_2$ , m), 3.99-4.05 (1H,  $\text{OCH}_2$ ,  $\text{H}_b$ , m), 4.17 (1H, Ar- $\text{OCH}_2$ ,  $\text{H}_a$ , dd,  $J$  4.9 and 10.1), 4.23 (1H, Ar $\text{OCH}_2$ ,  $\text{H}_b$ , dd,  $J$  5.2 and 10.1), 4.38 (2H,  $\text{OCH}_2$ , q,  $J$   
25 7.1), 4.89 (1H, CH- $\text{N}_3$ , t,  $J$  5.1), 7.13 (1H, Ar-H, dd,  $J$  2.1 and 8.4), 7.36 (1H, Ar-H, t,  $J$  7.9), 7.60 (1H, Ar-H, dd,  $J$  1.0 and 2.5) and 7.70 (1H, Ar-H, d,  $J$  7.8).

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- 45 -

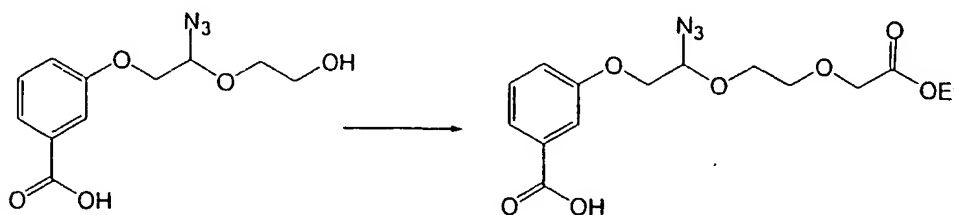
## 3-[2-Azido-2-(2-hydroxy-ethoxy)-ethoxy]-benzoic acid



5 3-[2-Azido-2-(2-hydroxy-ethoxy)-ethoxy]-benzoic acid  
ethyl ester (1.34 g, 4.55 mmol) was stirred with 4 M  
aqueous sodium hydroxide (11.4 ml, 45.5 mmol) and  
ethanol (11.4 ml) at room temperature. After 3 hrs,  
all the solvents were removed under reduced pressure  
10 and the residue was dissolved in 50 ml water. The  
solution was acidified with 1 N HCl to pH 2 and then  
extracted with DCM (3 x 50 ml). All the DCM extracts  
were combined, dried over MgSO<sub>4</sub> and evaporated under  
reduced pressure. The title compound was obtained as  
15 a colourless solid (1.2 g, 98.7%). ES-MS, m/z 265.85  
(M-1).

<sup>1</sup>HNMR [CDCl<sub>3</sub>]: 3.75-3.90 (3H, OCH<sub>2</sub>, H<sub>a</sub>, OCH<sub>2</sub>, m), 4.00-  
4.08 (1H, OCH<sub>2</sub>, H<sub>b</sub>, m), 4.17 (1H, Ar-OCH<sub>2</sub>, H<sub>a</sub>, dd, *J*  
20 4.8 and 10.1), 4.24 (1H, ArOCH<sub>2</sub>, H<sub>b</sub>, dd, *J* 5.1 and  
10.1), 4.90 (1H, CH-N<sub>3</sub>, t, *J* 5.1), 7.19 (1H, Ar-H, dd,  
*J* 2.5 and 8.2), 7.40 (1H, Ar-H, t, *J* 8.0), 7.60 (1H,  
Ar-H, s) and 7.70 (1H, Ar-H, d, *J* 7.9).

25 3-[2-Azido-2-(2-ethoxycarbonylmethoxy)-ethoxy]-  
benzoic acid

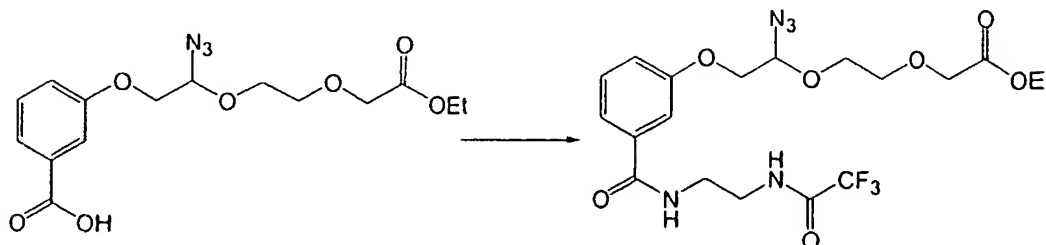


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To a solution of 3-[2-azido-2-(2-hydroxy-ethoxy)-ethoxy]-benzoic acid (0.535 g, 2 mmol) in dry THF (6 ml) was added NaH (60% dispersion, 0.246 g, 6 mmol) at 0 °C. After 10 minutes, ethyl-2-bromoacetate (0.488 ml, 4.4 mmol) was added. The reaction was then warmed up to room temperature and stirred for 4 hours. The reaction was quenched by pouring it into ice-cold water (50 ml). The mixture was extracted with DCM (2 x 50 ml) and the DCM extracts were discarded. The aqueous layer was then acidified to pH 2 with 1 N HCl, and extracted with DCM (2 x 50 ml). These DCM extracts were combined, dried over MgSO<sub>4</sub> and evaporated under reduced pressure. The residue was purified by column chromatography (1 x 20 cm). The title compound, eluted with 2% methanol in DCM, was obtained as an oil (0.223 g, 31.6%). ES-MS, m/z 351.95 (M-1).

<sup>1</sup>HNMR [CDCl<sub>3</sub>]: 1.29 (3H, CH<sub>3</sub>, t, J 7.2), 3.81 (2H, OCH<sub>2</sub>, t, J 4.4), 3.90 (1H, OCH<sub>2</sub>, H<sub>a</sub>, m), 4.04 (1H, OCH<sub>2</sub>, H<sub>b</sub>, m), 4.13-4.27 (6H, ArOCH<sub>2</sub>, OCH<sub>2</sub> and OCH<sub>2</sub>C(O)), 4.95 (1H, CH-N<sub>3</sub>, m), 7.19 (1H, Ar-H, dd, J 1.7 and 8.3), 7.40 (1H, Ar-H, t, J 7.8), 7.63 (1H, Ar-H, s) and 7.76 (1H, Ar-H, d, J 7.6).

[2-(1-Azido-2-{3-[2-(2,2,2-trifluoroacetyl-amino)-ethylcarbamoyl]-phenoxy}-ethoxy)-ethoxy]-acetic acid ethyl ester





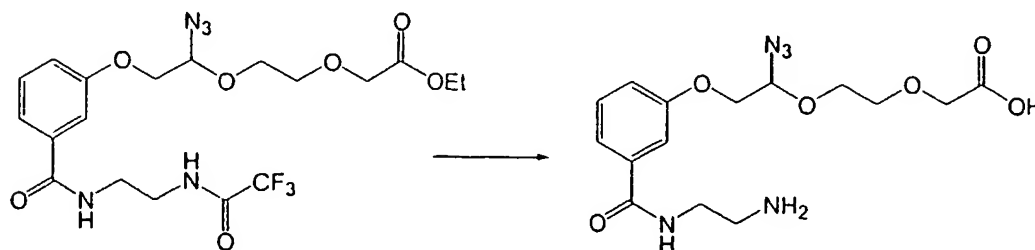
- 47 -

3-[2-Azido-2-(2-ethoxycarbonylmethoxy-ethoxy)-ethoxy]-benzoic acid (0.212 g, 0.6 mmol) was stirred with *N,N'*-disuccinimidyl carbonate (0.184 g, 0.72 mmol) and DMAP (0.088g, 0.72 mmol) in dry DMF (1 ml) at room temperature. After 10 minutes, trifluoroacetic acid salt of *N*-(2-amino-ethyl)-2,2,2-trifluoro-acetamide (0.194 g, 0.72 mmol) was added followed by diisopropylethylamine (DIPEA) (0.251 ml, 1.44 mmol). The reaction mixture was then stirred at room temperature for 17 hours. All the solvents were evaporated under reduced pressure and the residues were partitioned between DCM (50 ml) and aqueous  $\text{NaH}_2\text{PO}_4$  (1 N, 50 ml). The aqueous layer was further extracted with DCM (2 x 25 ml). All the DCM extracts were combined, dried over  $\text{MgSO}_4$  and evaporated under reduced pressure. The residue was purified by column chromatography (1 x 20 cm). The title compound, eluted with 1 % methanol in DCM, was obtained as an oil (0.255 g, 86.6%). ES-MS,  $m/z$  490.10 ( $M-1$ ).

$^1\text{H}$ NMR [ $\text{CDCl}_3$ ]: 1.28 (3H,  $\text{CH}_3$ , t,  $J$  7.1), 3.60 (2H,  $\text{CH}_2\text{N}$ , m), 3.69 (2H,  $\text{CH}_2\text{N}$ , m), 3.80 (2H,  $\text{OCH}_2$ , t,  $J$  4.3), 3.86 (1H,  $\text{OCH}_2$ ,  $\text{H}_a$ , m), 4.04 (1H,  $\text{OCH}_2$ ,  $\text{H}_b$ , m), 4.10-4.25 (6H,  $\text{ArOCH}_2$ ,  $\text{OCH}_2$  and  $\text{OCH}_2\text{C(O)}$ , m), 4.92 (1H,  $\text{CH-N}_3$ , m), 6.85 (1H, NH, br), 7.09 (1H, Ar-H, m), 7.37 (3H, Ar-H, m), and 7.96 (1H, NH, br).

(2-{2-[3-(2-Amino-ethylcarbamoyl)-phenoxy]-1-azido-ethoxy}-ethoxy)-acetic acid

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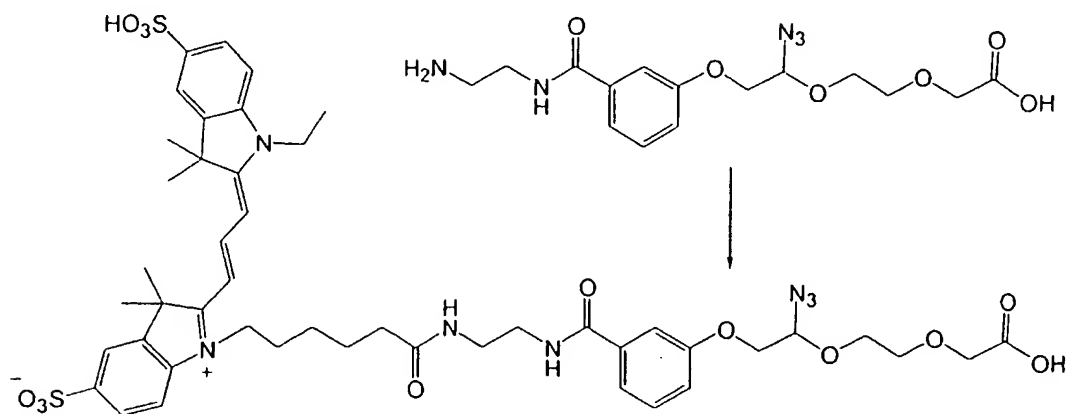
- 48 -

[2-(1-Azido-2-{3-[2-(2,2,2-trifluoroacetyl-amino)-ethoxycarbonyl]-phenoxy}-ethoxy)-ethoxy]-acetic acid ethyl ester (0.196 g, 0.4 mmol) was stirred with 4 M aqueous sodium hydroxide (1 ml, 4 mmol) and ethanol (1 ml) at room temperature. After 2 hrs, all the solvents were removed under reduced pressure and the residue was dissolved in 15 ml water. The solution was extracted with DCM (2 x 15 ml). The DCM extracts were discarded and the aqueous layer was acidified with 1 N HCl to pH 2. Then the solution was extracted again with DCM (3 x 15 ml). The DCM extracts were discarded and the aqueous layer was neutralised with 1 N NaOH to pH 8 and then evaporated under reduced pressure to dryness. The white solids were triturated with DCM/MeOH (v/v; 1:1, 2 x 25 ml). All the solids were filtered off and the filtrates were combined and evaporated under reduced pressure to give a gum. The gum was added in 10% MeOH in DCM (15 ml) and the insoluble, white solids were filtered off. The filtrates were evaporated under reduced pressure to give the mono-sodium salt of the title compound as a colourless powder (0.135 g, 86.6 %). ES-MS, m/z 368.00 (M+1).

<sup>1</sup>HNMR [D<sub>2</sub>O]: 3.01 (2H, CH<sub>2</sub>NH<sub>2</sub>, t, J 6.0), 3.51 (2H, CH<sub>2</sub>N, t, J 6.0), 3.62 (2H, OCH<sub>2</sub>, m), 3.77 (1H, OCH<sub>2</sub>, H<sub>a</sub>, m), 3.80 (2H, CH<sub>2</sub>C(O), s), 3.96 (1H, OCH<sub>2</sub>, H<sub>b</sub>, m), 4.19 (2H, ArOCH<sub>2</sub>, d, J 4.3), 5.01 (1H, CH-N<sub>3</sub>, t, J 4.5), 7.13 (1H, Ar-H, d, J 7.9) and 7.25-7.39 (3H, Ar-H, m).

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[2- (1-Azido-2- {3- [2- (6-Cy3-hexanoylamino) -  
ethylcarbamoyl] -phenoxy} -ethoxy) -ethoxy] -acetic acid



5

The commercial Cy3 mono *N*-hydroxysuccinimide ester (5 mg, 6.53  $\mu$ mol) and (2- {2- [3- (2-amino-ethylcarbamoyl) -phenoxy] -1-azido-ethoxy} -ethoxy) -acetic acid (7.2 mg, 19.6  $\mu$ mol) were stirred together in dry DMF. DIPEA  
 10 (6.82  $\mu$ l, 39.2  $\mu$ mol) was added. After 2 hr stirring at room temperature, all the solvent was evaporated under reduced pressure. The residue was purified by preparative HPLC [HPLC gradient: A, 100% 0.1 M TEAB; B 100% MeCN; 0-2 min, 5% B (flow 2-10 ml/min); 2-19  
 15 min, 5-45% B (flow 10 ml/min); 19-21 min, 45-95% B (flow 10 ml/min); 21-24 min, 95% B (flow 10 ml/min); 24-26 min, 95-5% B (flow 10 ml/min); 26-30 min, 5% B (flow 10-2 ml/min)]. The title compound with retention time of 17.85 min was obtained as a pink  
 20 solid (4.93  $\mu$ mol, 75.5%, quantification at 550 nm in water). ES-MS, *m/z* 488.95 [(*M*/2)-1].  $^1\text{H}$ NMR in  $\text{D}_2\text{O}$  indicated approximately 1.8 triethylammonium count ions.

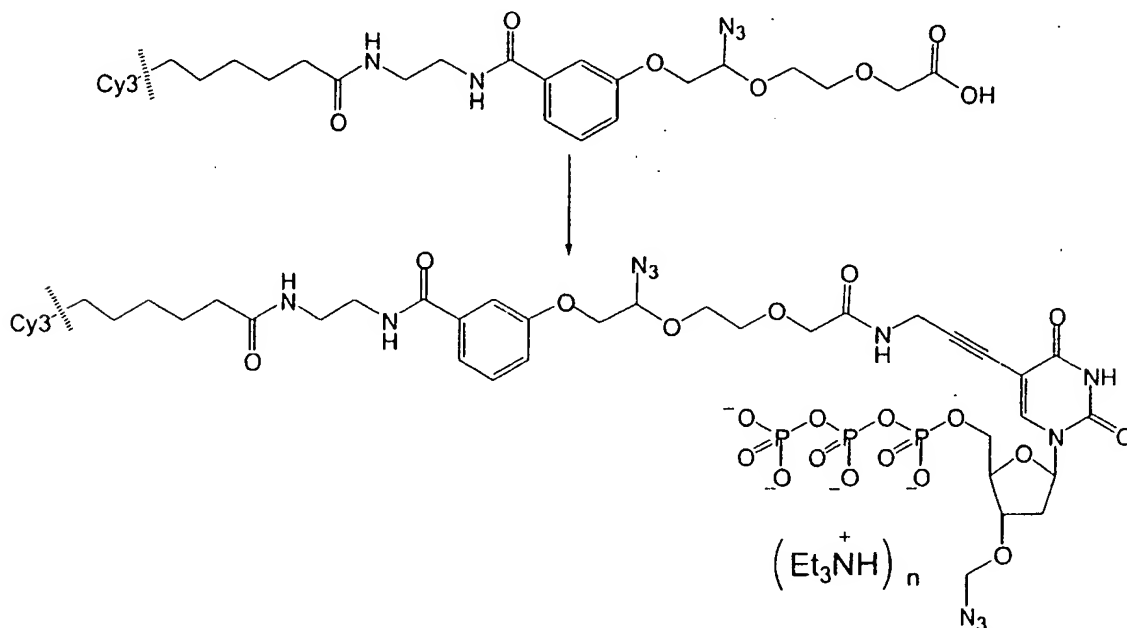
25  $^1\text{H}$ NMR [ $\text{D}_2\text{O}$ ]: 1.15 (16.2H,  $\text{CH}_3$  ( $\text{Et}_3\text{N}$ ), t, *J* 7.2), 1.21 (3H,  $\text{CH}_3$ , t, *J* 7.1), 1.47 (2H,  $\text{CH}_2$ , m), 1.55 (2H,  $\text{CH}_2$ , m), 1.58 (6H, 2 x  $\text{CH}_3$ , s), 1.61 (6H, 2 x  $\text{CH}_3$ , s), 2.10 (2H,  $\text{CH}_2\text{C}(\text{O})$ , t, *J* 6.5), 3.06 (10.8 H,  $\text{CH}_2$  ( $\text{Et}_3\text{N}$ ), q, *J*

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7.2), 3.23 (2H, CH<sub>2</sub>N, t, *J* 5.5), 3.32 (2H, CH<sub>2</sub>N, t, *J* 5.8), 3.56 (2H, OCH<sub>2</sub>, m), 3.67-3.78 (3H, OCH<sub>2</sub>, H<sub>a</sub> and CH<sub>2</sub>N, m), 3.79 (2H, OCH<sub>2</sub>C(O), s), 3.85-3.97 (3H, OCH<sub>2</sub>, H<sub>b</sub> and CH<sub>2</sub>N, m), 3.98 (2H, ArOCH<sub>2</sub>, d, *J* 4.4), 4.85  
 5 (1H, CH-N<sub>3</sub>, t, *J* 4.3), 6.14 (1H, =CH, d, *J* 13.4), 6.19 (1H, =CH, d, *J* 13.4), 6.90 (1H, Ar-H, m), 7.10-7.19 (5H, Ar-H, m), 7.69 (2H, Ar-H, d, *J* 8.4), 7.73 (1H, Ar-H, s), 7.77 (1H, Ar-H, s) and 8.36 (1H, =CH, t, *J* 13.4).

10

**5-[3-(-Cy3-azidolinkeracetyl-amino)-prop-1-ynyl]-3'-azidomethoxy-dUTP**



15 [2-(1-Azido-2-{3-[2-(6-Cy3-hexanoylamino)-ethylcarbamoyl]-phenoxy}-ethoxy)-ethoxy]-acetic acid (2 μmol) was stirred with *N,N'*-disuccinimidyl carbonate (0.563 mg, 2.2 μmol) and DMAP (0.269 mg, 2.2 μmol) in dry DMF (1 ml) at room temperature. After  
 20 10 minutes, all the reaction mixture was added to a solution of the tri-*n*-butyl ammonium salt of [5-(3-amino-prop-1-ynyl)]-3'-azidomethoxy-dUTP (6 μmol, prepared by evaporating an aqueous solution of [5-(3-amino-prop-1-ynyl)]-3'-azidomethoxy-dUTP with tri-*n*-

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butyl amine (72  $\mu$ l, 300  $\mu$ mol)). The reaction mixture was sonicated for 5 minutes, then stirred at room temperature for 3hrs. The reaction mixture was diluted with chilled 0.1 M TEAB (10 ml), the resulting solution was applied onto a short column of DEAE A-25 (1 x 10 cm). The column was initially eluted with 0.1 M TEAB buffer (50 ml) and then 1.0 M buffer (50 ml). The 1.0 M TEAB eluates were collected and evaporated under reduced pressure. The residue was co-evaporated with MeOH (2 x 10 ml) and then purified by semi-preparative HPLC [HPLC gradient: A, 100% 0.1 M TEAB; B 100% MeCN; 0-2 min, 5% B (flow 2-5 ml/min); 2-14 min, 5-20% B (flow 5 ml/min); 14-20 min, 20-23% B (flow 5 ml/min); 20-22 min, 23-95% B (flow 5 ml/min); 22-25 min, 95% B (flow 5 ml/min); 25-26 min, 95-5% B (flow 5 ml/min); 26-30 min, 5% B (flow 5-2 ml/min)]. The product with retention time of 19.2 min was collected and evaporated under reduced pressure and the residue was co-evaporated with methanol (3 x 5 ml) to give the title compound as triethyl ammonium salt (1.22  $\mu$ mol, quantification at  $\lambda_{550}$  in 0.1 M TEAB buffer, 61.1%).  $^1\text{H}$ NMR in  $\text{D}_2\text{O}$  indicated approximately seven triethylammonium count ions.

$^1\text{H}$ NMR [ $\text{D}_2\text{O}$ ]: 1.14 (63H,  $\text{CH}_3$  ( $\text{Et}_3\text{N}$ ), t,  $J$  7.3), 1.23 (3H,  $\text{CH}_3$ , t,  $J$  7.1), 1.49 (2H,  $\text{CH}_2$ , m), 1.55 (2H,  $\text{CH}_2$ , m), 1.60 (6H, 2 x  $\text{CH}_3$ , s), 1.62 (6H, 2 x  $\text{CH}_3$ , s), 1.95-2.07 (1H,  $\text{H}_a\text{-2'}$ , m), 2.13 (2H,  $\text{CH}_2\text{C(O)}$ , t,  $J$  6.7), 2.17-2.27 (1H,  $\text{H}_b\text{-2'}$ , m), 3.05 (42 H,  $\text{CH}_2$  ( $\text{Et}_3\text{N}$ ), q,  $J$  7.3), 3.27 (2H,  $\text{CH}_2\text{N}$ , t,  $J$  5.7), 3.37 (2H,  $\text{CH}_2\text{N}$ , t,  $J$  5.8), 3.55-3.79 (5H, m), 3.80-4.10 (11H, m), 4.15 (1H,  $\text{H-4'}$ , m), 4.38 (1H,  $\text{H-3'}$ , m), 4.80 (2H,  $\text{OCH}_2\text{-N}_3$ , s), 4.88 (1H,  $\text{CH-N}_3$ , m), 5.95 (1H,  $\text{H-1'}$ , q,  $J$  7.6), 6.13 (1H,  $=\text{CH}$ , d,  $J$  13.4), 6.20 (1H,  $=\text{CH}$ , d,  $J$  13.4), 6.84 (1H,  $\text{Ar-H}$ , d,  $J$  7.3), 7.05-7.24 (5H,  $\text{Ar-H}$ , m), 7.60-7.80

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(5H, Ar-H and H-6, m) and 8.35 (1H, =CH, t,  $J$  13.4).

$^{31}\text{P}$ NMR [ $\text{D}_2\text{O}$ ]: -20.82 ( $^{\beta}\text{P}$ , m), -10.07 ( $^{\alpha}\text{P}$ , d,  $J$  16.2) and -4.90 ( $^{\gamma}\text{P}$ , d,  $J$  18.9).

5     **2 cycles of fully functional nucleoside triphosphate (FFN)**

**Preparation of beads**

Take 15  $\mu\text{L}$  of Dynabeads<sup>®</sup> M-280 streptavidin coated  
10     beads (DynaL Biotech), remove storage buffer and wash  
3 times with 150  $\mu\text{L}$  of TE buffer (Tris.HCl pH 8, 10 mM  
and EDTA, 1 mM). Resuspend in 37.5  $\mu\text{L}$  of B & W  
buffer (10 mM Tris-HCl pH 7.5, 1 mM EDTA and 2.0 M  
NaCl), add 10  $\mu\text{L}$  of biotinylated  $^{32}\text{P}$  labelled hairpin  
15     DNA and add 27.5  $\mu\text{L}$  of water. Allow to stand at room  
temperature for 15 minutes. Remove buffer and wash  
beads 3 times with 100  $\mu\text{L}$  of TE buffer.

**Incorporation of the 1<sup>st</sup> FFN**

20     75  $\mu\text{L}$  reaction, Tris.HCl pH 8.8 50 mM, Tween-20,  
0.01%,  $\text{MgSO}_4$  4 mM,  $\text{MnCl}_2$ , 0.2 mM, add 2  $\mu\text{M}$  FFN and 100  
nM polymerase (*Thermococcus* sp. 9<sup>°</sup>N exo Y409V A485L  
supplied by New England Biolabs). This solution is  
then added to the beads and mixed thoroughly and  
25     incubated at 65°C taking 5  $\mu\text{L}$  samples at 3 and 10  
minutes and stopping with 5  $\mu\text{L}$  of gel loading buffer  
(xylene cyanol - bromophenol blue dye solution, Sigma-  
Aldrich). The reaction mixture is removed from the  
remaining beads and the beads washed 3 times with 100  
30      $\mu\text{L}$  of TE buffer.

**Chase step**

A sample was removed from the incorporation reaction  
and added to 1  $\mu\text{M}$  of dNTPs (0.25  $\mu\text{M}$  each). This was

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stopped after 10 minutes by adding 5  $\mu$ L of gel loading buffer.

#### Deblocking step

5 50  $\mu$ L of Tris-(2-carboxyethyl)phosphine, trisodium salt (TCEP) 0.1M is added to the beads and mixed thoroughly. The mixture was then incubated at 65°C for 15 minutes. The deblocking solution is removed and the beads washed 3 times with 100  $\mu$ L TE buffer.  
10 The beads were then resuspended in 50  $\mu$ L of TE and a 5  $\mu$ L sample was removed and 5  $\mu$ L of gel loading buffer)

#### Incorporation of the 2<sup>nd</sup> FFN

20  $\mu$ L reaction, Tris.HCl pH 8.8 50 mM, Tween-20, 0.01%, MgSO<sub>4</sub> 4 mM, MnCl<sub>2</sub> 0.4 mM, add 2  $\mu$ M FFN and 100  
15 nM polymerase (*Thermococcus* sp. 9°N exo Y409V A485L supplied by New England Biolabs). This solution is then added to the beads and mixed thoroughly and incubated at 65°C taking 5 $\mu$ L samples at 3 and 10  
20 minutes and stopping with 5  $\mu$ L of gel loading buffer.

#### Chase step

A sample was removed from the incorporation reaction and added to 1 $\mu$ M of dNTPs (0.25  $\mu$ M each). This was  
25 stopped after 10 minutes by adding 5  $\mu$ L of gel loading buffer.

Before loading the samples onto a denaturing 12% acrylamide sequencing gel 0.5  $\mu$ L of EDTA (0.5 M) was  
30 added to each sample and then heated to 95 °C for 10 minutes.

All patents, patent applications, and published references cited herein are hereby incorporated by

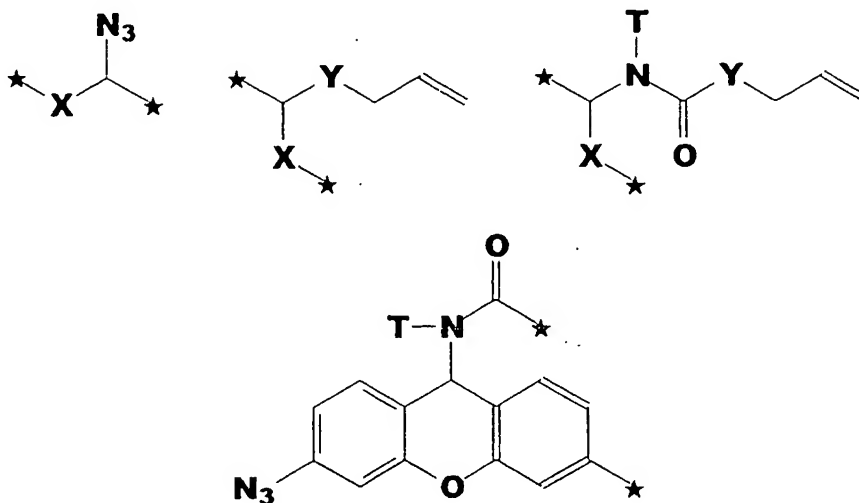
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reference in their entirety. While this invention has  
been particularly shown and described with references  
to preferred embodiments, it will be understood by  
those skilled in the art that various changes in form  
5 and details may be made without departing from the  
scope of the invention encompassed by the claims.



## CLAIMS

1. A nucleotide or nucleoside having a base attached to a detectable label via a cleavable linker,  
 5 characterised in that the cleavable linker contains a moiety selected from the group comprising:



10

- (wherein X is selected from the group comprising O, S, NH and NQ wherein Q is a C<sub>1-10</sub> substituted or unsubstituted alkyl group, Y is selected from the group comprising O, S, NH and N(allyl), T is hydrogen or a C<sub>1-10</sub> substituted or unsubstituted alkyl group and  
 15 \* indicates where the moiety is connected to the remainder of the nucleotide or nucleoside).

2. The nucleotide or nucleoside as claimed in claim 1 wherein X is O or S.  
 20

3. The nucleotide or nucleoside as claimed in claim 1 or claim 2 wherein Y is O or S.

- 25 4. The nucleotide or nucleoside as claimed in any one of claims 1 to 3 wherein Y is O.

5. The nucleotide or nucleoside as claimed in any

one preceding claim wherein the moiety may be present in the nucleotide or nucleoside in either of two orientations.

5       6.    The nucleotide or nucleoside as claimed in any one preceding claim wherein the base is a purine, or a pyrimidine.

10       7.    The nucleotide or nucleoside as claimed in any one preceding claim wherein the linker is attached to the 5-position of a pyrimidine or 7-position of a purine.

15       8.    The nucleotide or nucleoside as claimed in any one preceding claim wherein the base is a deazapurine.

20       9.    The nucleotide or nucleoside as claimed in any one preceding claim wherein the nucleotide has a ribose or deoxyribose sugar moiety.

25       10.   The nucleotide or nucleoside as claimed in claim 9 wherein the ribose or deoxyribose sugar comprises a hydroxyl protecting group attached to the 2' or 3' oxygen atom.

30       11.   The nucleotide or nucleoside as claimed in claim 10 wherein the same chemical conditions may be used to effect cleavage of the cleavable linker and to remove the hydroxyl protecting group.

35       12.   The nucleotide or nucleoside as claimed in any one preceding claim wherein the nucleotide is a deoxyribonucleotide triphosphate.

13.   The nucleotide or nucleoside as claimed in any

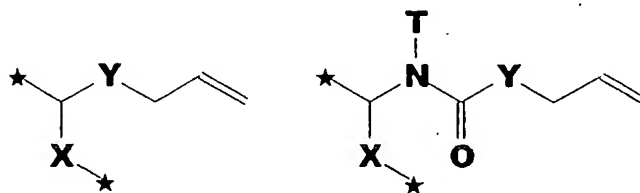
one preceding claim wherein the detectable label is a fluorophore.

14. An oligonucleotide comprising one or more  
5 nucleotides as defined in any one of claims 1 to 13.

15. The oligonucleotide as claimed in claim 14 wherein at least one nucleotide is present at a terminal position in said oligonucleotide.

10

16. A method of cleaving a linker that contains a moiety selected from the groups comprising:



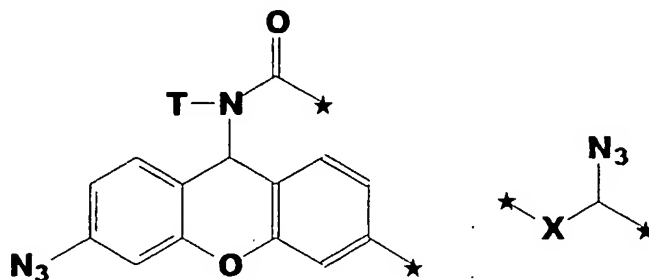
15

(wherein X is selected from the group comprising O, S, NH and NQ wherein Q is a C<sub>1-10</sub> substituted or unsubstituted alkyl group, Y is selected from the group comprising O, S, NH and N(allyl), T is hydrogen or a C<sub>1-10</sub> substituted or unsubstituted alkyl group and  
20 \* indicates where the moiety is connected to the remainder of a nucleotide or nucleoside),  
said linker being present in a nucleotide or nucleoside and connecting the base thereof to a  
25 detectable label, said method comprising contacting the nucleotide or nucleoside with a water-soluble phosphine-based transition metal catalyst.

17. The method as claimed in claim 16 wherein the  
30 transition metal is selected from the group comprising platinum, palladium, rhodium, ruthenium, osmium and iridium.

18. The method as claimed in claim 16 wherein the transition metal is palladium.

19. A method of cleaving a linker that contains a moiety selected from the groups comprising:



(wherein X is selected from the group comprising O, S, NH and NQ wherein Q is a C<sub>1-10</sub> substituted or unsubstituted alkyl group, T is hydrogen or a C<sub>1-10</sub> substituted or unsubstituted alkyl group and \* indicates where the moiety is connected to the remainder of a nucleotide or nucleoside),

15        said linker being present in a nucleotide or nucleoside and connecting the base thereof to a detectable label, said method comprising contacting the nucleotide or nucleoside with a water-soluble phosphine.

20

20. The method as claimed in any one of claims 16 to 19 wherein said phosphine is a derivatised triaryl phosphine or a derivatised trialkyl phosphine.

25        21. The method as claimed in any one of claims 16 to 20 wherein said phosphine is a triaryl phosphine derivatised with one or more functionalities selected from the group comprising amino, hydroxyl, carboxyl and sulfonate.

30

22. The method as claimed in any one of claims 16 to

21 wherein the water-soluble phosphine is selected from the group comprising 3,3',3''-phosphinidynetris (benzenesulfonic acid) or *tris*(2-carboxyethyl)phosphine and their salts.

5

23. The method as claimed in any one of claims 16 to 19 wherein said phosphine contains one or more nitrogen atoms.

10

24. The method as claimed in any one of claims 16 to 23 wherein X is O or S.

25. The method as claimed in any one of claims 16 to 24 wherein Y is O or S.

15

26. The method as claimed in any one of claims 16 to 25 wherein Y is O.

20

27. The method as claimed in any one of claims 16 to 26 wherein the moieties may be present in the nucleoside or nucleotide in either of two orientations.

25

28. The method of in any one of claims 16 to 27 wherein said label is detected before said linker is cleaved.

30

29. The method of claim 28 wherein said method involves cleavage of the linker in a nucleotide which is incorporated into an oligonucleotide.

35

30. The method of claim 29 wherein said incorporated nucleotide is present at a terminal position in said oligonucleotide.

31. The method as claimed in any one of claims 16 to

30 wherein the base is a purine, or a pyrimidine.

32. The method of claim 31, wherein the linker is  
attached to the 5-position of a pyrimidine or 7-  
5 position of a purine.

33. The method as claimed in any one of claims 16 to  
32 wherein the base is a deazapurine.

10 34. The method as claimed in any one of claims 16 to  
33 wherein the nucleotide has a ribose or deoxyribose  
sugar moiety.

35. The method as claimed in claim 34 wherein the  
15 ribose or deoxyribose sugar comprises a hydroxyl  
protecting group attached to the 2' or 3' oxygen atom.

36. The method as claimed in any one of claims 16 to  
35 wherein the nucleotide is a deoxyribonucleotide  
20 triphosphate.

37. The method as claimed in any one of claims 16 to  
36 wherein the detectable label is a fluorophore.

25 38. The method as claimed in any one of claims 29 to  
37 wherein the incorporating step is effected by a  
reverse transcriptase, a terminal transferase or a  
polymerase.

30 39. The method of claim 38 wherein the polymerase is  
a *Thermococcus* sp.

40. The method of claim 39 wherein the *Thermococcus*  
sp is 9°N or a single mutant or double mutant thereof.

35

41. The method of claim 40 wherein the double mutant

is -Y409V A485L.

42. The method as claimed in any one of claims 29 to 41 wherein the detectable label and/or the cleavable  
5 linker is of a size sufficient to prevent the incorporation of a subsequent nucleotide into the nascent oligonucleotide.

43. The method as claimed in any one of claims 29 to  
10 42 wherein the incorporated nucleotide contains a 3'OH blocking group which serves to prevent incorporation of any further nucleotides.

44. The method as claimed in claim 43 wherein the  
15 same chemical conditions used to effect cleavage of the cleavable linker serve to remove the 3'OH blocking group.

45. The method as claimed in any one of claims 29 to  
20 44 wherein the detecting step permits the identification of the incorporated nucleotide.

46. A method for determining the identity of a nucleotide in a target single-stranded polynucleotide,  
25 comprising:

(a) providing one or more of the nucleotides A, G, C and T or U in which each of said nucleotides has a base that is attached to a distinct detectable label via a linker, said linker being cleavable with a  
30 water-soluble phosphine; and a nascent polynucleotide complementary to the target polynucleotide, one of said provided nucleotides being suitable for incorporation into said nascent polynucleotide;

(b) incorporating the nucleotide suitable for  
35 incorporation into said nascent polynucleotide; and

(c) carrying out a method as defined in claim 45.

47. The method as claimed in claim 46 wherein steps (a) and (b) are repeated one or more times so as to determine the identity of a plurality of bases in the target polynucleotide.

5

48. A method as claimed in claim 46 or claim 47 wherein step (a) comprises contacting the provided nucleotides with the target sequentially.

10

49. A method as claimed in any one of claims 46 to 48 wherein step (a) comprises at least one substep of providing one of the four said nucleotides.

15

50. A method as claimed in claim 49 wherein step (a) further comprises, after said substep, providing the other three nucleotides simultaneously or sequentially.

20

51. A method as claimed in claim 50 wherein said other three nucleotides are added sequentially, either by providing them one at a time; or two simultaneously and then the remaining one; or one of the three and then the remaining two simultaneously.

25

52. A method as claimed in any one of claims 46 to 48 wherein step (a) comprises at least a substep of providing two of the four said nucleotides.

30

53. A method as claimed in claim 52 wherein step (a) further comprises, after said substep, providing the other two nucleotides simultaneously or sequentially.

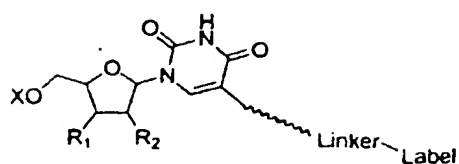
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54. A method as claimed in any one of claims 46 to 48 wherein step (a) comprises at least a substep of providing three of the four said nucleotides.

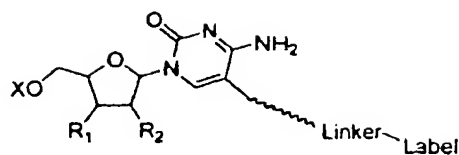


55. A method as claimed in claim 54 wherein step (a) further comprises, after said substep, providing the remaining nucleotide of the four said nucleotides.
- 5 56. A method as claimed in any one of claims 46 to 48 wherein step (a) comprises providing all four of the said nucleotides and contacting them with the target simultaneously.
- 10 57. A method as claimed in any one of claims 46 to 56 wherein any unincorporated nucleotides are removed prior to the provision of further nucleotide(s) and/or the effecting of step (c).
- 15 58. A method as claimed in claim 57 wherein step (c) is effected without unsuitable nucleotides having been provided after provision of said suitable nucleotide.
59. Use of a nucleotide as defined in any one of  
20 claims 1 to 13 in a Sanger or Sanger-type sequencing method.
60. A method of using a nucleotide of claims 1  
25 wherein said method includes a Sanger or Sanger-type sequencing method.

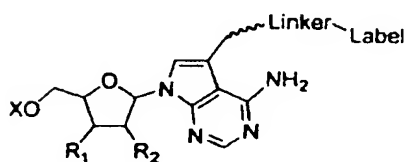
1/4



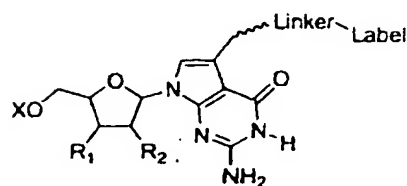
Uridine C5-linker



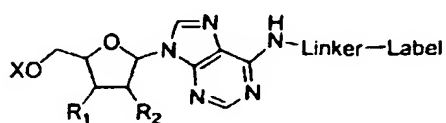
Cytidine C5-linker



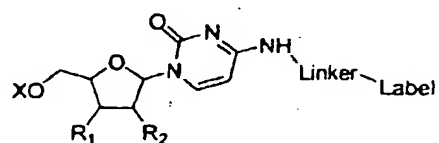
N7 Deazaadenosine C7-linker



N7 Deazaguanosine C7-linker



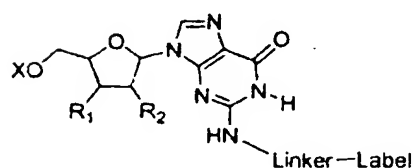
Adenosine N6-linker



Cytidine N4-linker

where  $R_1$  and  $R_2$ , which may be the same or different, are each selected from H, OH, or any group which can be transformed into an OH. Suitable groups for  $R_1$  and  $R_2$  are described in Figure 3

X = H, phosphate, diphosphate or triphosphate



Guanosine N2-linker

Fig. 1

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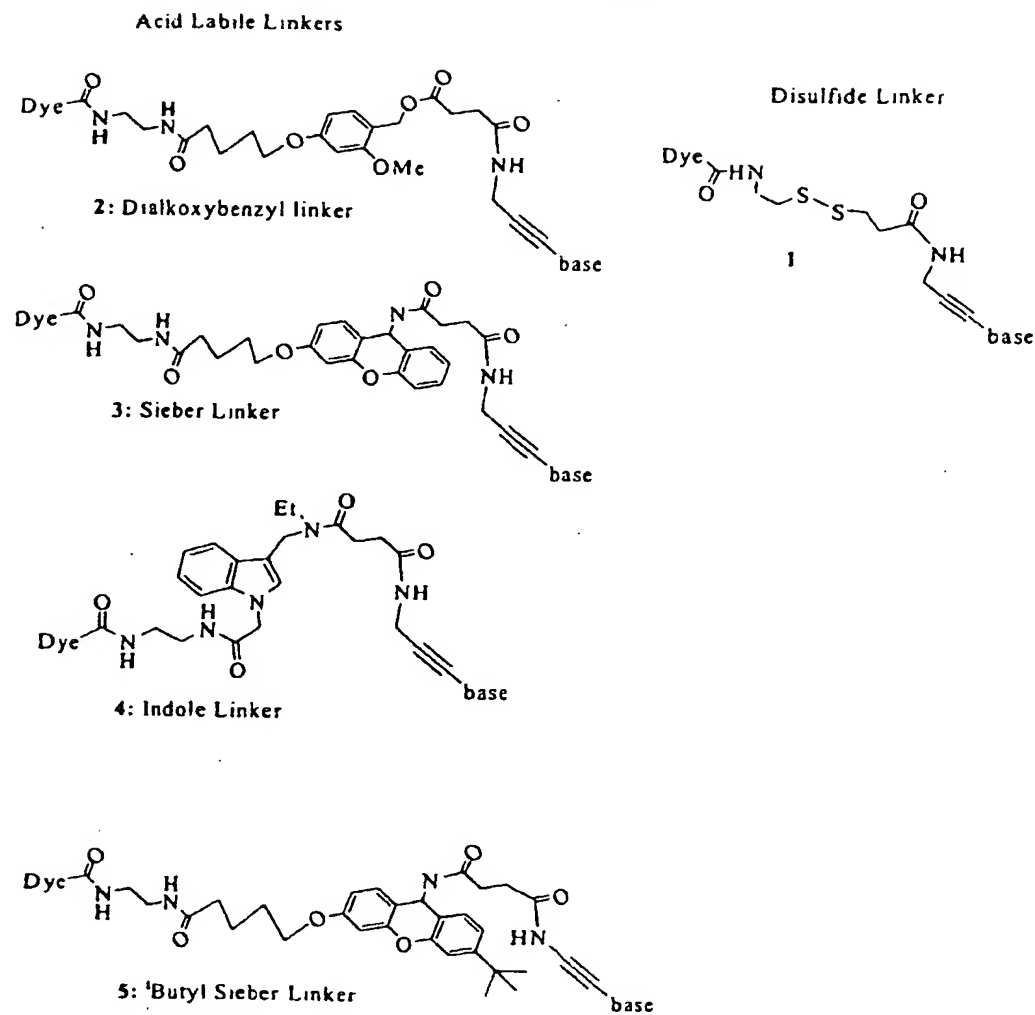
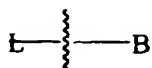
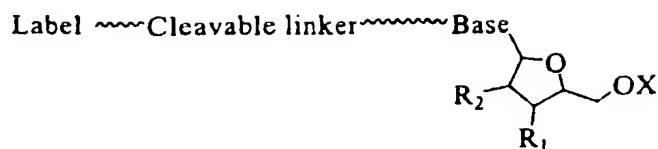


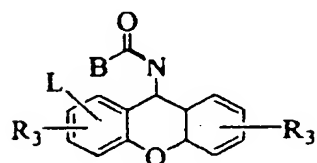
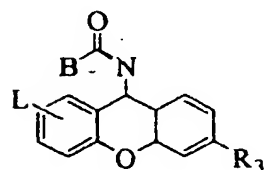
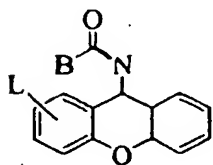
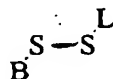
Fig. 2

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Cleavable linkers may include:

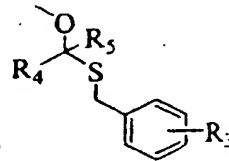
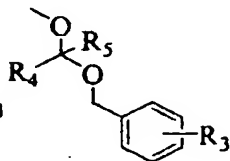
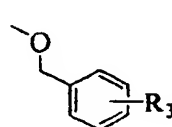
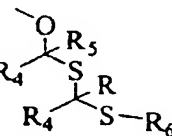
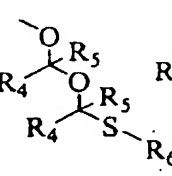
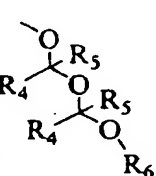
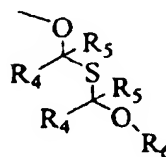
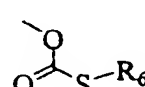
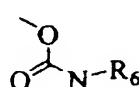
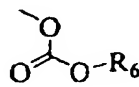
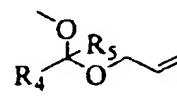
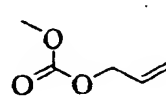
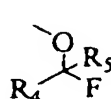
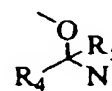
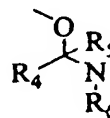
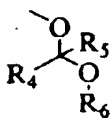
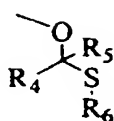


R3 represents one or more substituents independently selected from alkyl, alkoxy, amino or halogen

Alternatively, cleavable linkers may be constructed from any labile functionality used on the 3'-block

where  $R_1$  and  $R_2$ , which may be the same or different, are each selected from H, OH, or any group which can be transformed into an OH, including a carbonyl

$R_1$  and  $R_2$  groups may include



where  $R_4$  is H or alkyl,  $R_5$  is H or alkyl and  $R_6$  is alkyl, cycloalkyl, alkenyl, cycloalkenyl or benzyl

and X is H, phosphate, diphosphate or triphosphate

Fig. 3

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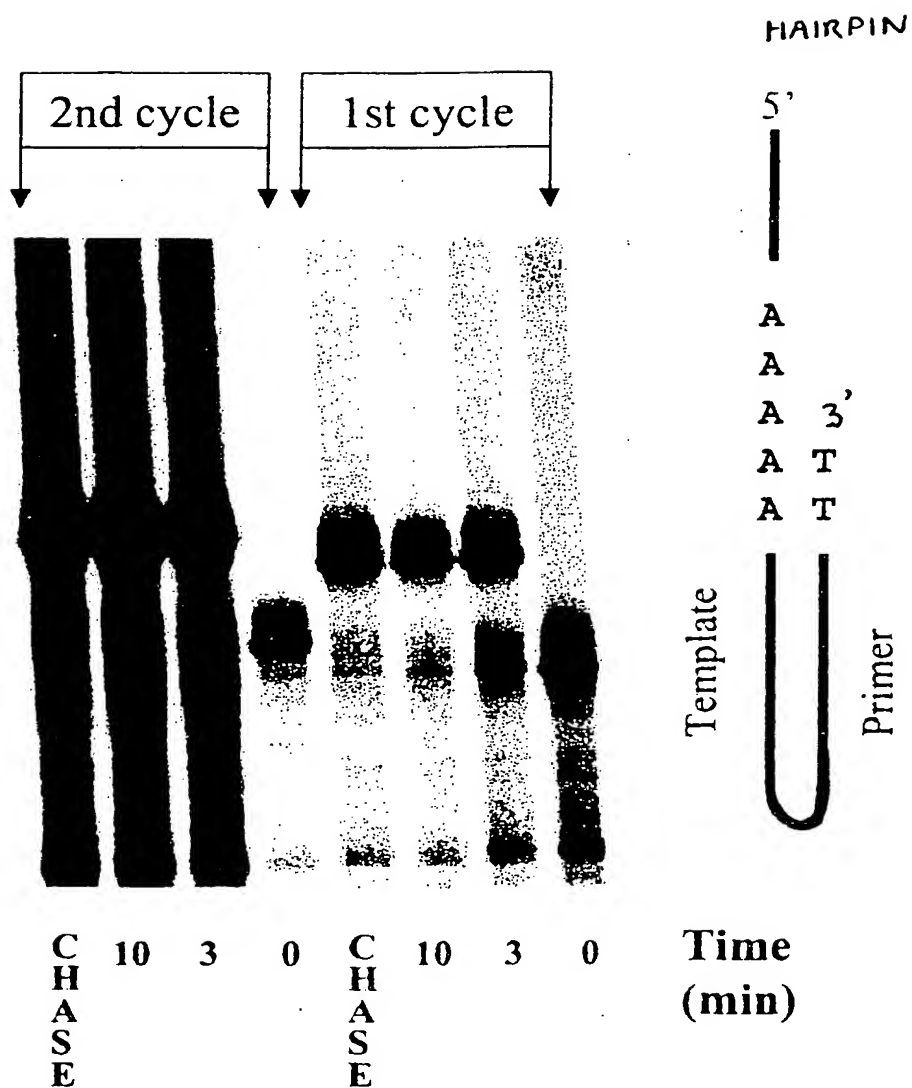


Fig. 4

# INTERNATIONAL SEARCH REPORT

Intern .pplication No  
PCT/GB J3/03690

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C07H19/06 C07H19/10 C07H19/16 C07H19/20 C07H19/23  
C07H21/00 C12Q1/68 G01N33/53

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C07H C12Q G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, CHEM ABS Data

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 99 57321 A (REHA KRANTZ LINDA J ;UNIV ALBERTA (CA); PIZZICONI VINCENT B (US);) 11 November 1999 (1999-11-11) the whole document	1,46
A	WO 02 29003 A (UNIV COLUMBIA) 11 April 2002 (2002-04-11) cited in the application the whole document	1,46

☐ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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- \*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- \*&\* document member of the same patent family

Date of the actual completion of the international search

7 January 2004

Date of mailing of the international search report

16/01/2004

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2  
NL - 2280 HV Rijswijk  
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,  
Fax: (+31-70) 340-3016

Authorized officer

de Nooy, A

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Internal — Application No

PCT/GB 03/03690

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